



Hot Line Page #	Test Number	Summary of Changes by Test Name	Name Change	Methodology	Performed/Reported Schedule	Specimen Requirements	Reference Interval	Interpretive Data	Note	CPT Code	Component Change	Other Interface Change	New Test	Inactive
6	0060152	Acid-Fast Bacillus (AFB) Culture and AFB Stain				х			х					
6	0020008	Alanine Aminotransferase, Serum or Plasma										x		
6	<u>2005736</u>	Alkaline Phosphatase Isoenzymes, CSF			х									
6	0021020	Alkaline Phosphatase Isoenzymes, Serum or Plasma				х								
7	<u>0060217</u>	Antimicrobial Susceptibility, AFB/Mycobacteria					х							
48	<u>0055566</u>	Apolipoprotein E (<i>APOE</i>) 2 Mutations, Cardiovascular Risk												x
10	<u>2013341</u>	Apolipoprotein E (<i>APOE</i>) Genotyping, Alzheimer Disease Risk											x	
11	<u>2013337</u>	Apolipoprotein E (<i>APOE</i>) Genotyping, Cardiovascular Risk											x	



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12	<u>2013327</u>	Aquaporin-4 Receptor Antibody by ELISA with Reflex to Aquaporin-4 Receptor Antibody, IgG by IFA											x	
13	<u>2013320</u>	Aquaporin-4 Receptor Antibody, IgG by IFA with Reflex to Titer, Serum											x	
13	<u>0095227</u>	Arylsulfatase A, 24-Hour Urine		х	х						х			
13	0020007	Aspartate Aminotransferase, Serum or Plasma										х		
14	2008901	B-Cell Memory and Naive Panel					х	х	х	х	х			
15	<u>2008420</u>	<i>BCR-ABL1</i> Mutation Analysis for Tyrosine Kinase Inhibitor Resistance by Next Generation Sequencing							x					
15	<u>2005017</u>	BCR-ABL1, Major (p210), Quantitative									х			
15	<u>2005010</u>	<i>BCR-ABL1</i> , Qualitative with Reflex to <i>BCR-ABL1</i> Quantitative									x			
48	<u>0055691</u>	BIRC2-MALT1 (<i>API2-MALT1</i>) Translocation, t(11;18) by RT-PCR												x
48	<u>0051434</u>	Bloom Syndrome (BLM) 1 Mutation, Fetal												x
48	<u>0051454</u>	Canavan Disease (ASPA) 4 Mutations, Fetal												х
15	<u>2011450</u>	Carisoprodol and Meprobamate, Serum or Plasma, Quantitative					x					x		
48	0021021	Carotenes, Fractionated, Plasma or Serum												x
16	0040002	CBC with Platelet Count				х	х				х			
17	0040003	CBC with Platelet Count and Automated Differential				х	х				х			
18	2012717	CHARGE Syndrome (CHD7) Sequencing, Fetal				х								
18	<u>0060850</u>	Chlamydia trachomatis Culture			х	х			х					
18	<u>0080469</u>	Chromogranin A						х						
19	0060851	Clostridium difficile Cytotoxin Cell Assay			х									
19	<u>2013259</u>	Comprehensive Kidney Biopsy Workup											X	
19	<u>0020408</u>	Comprehensive Metabolic Panel										x		
20	<u>2013260</u>	Comprehensive Muscle Biopsy Workup											х	
21	<u>2013261</u>	Comprehensive Nerve Biopsy Workup											х	
22	<u>2013257</u>	Consultation, Head and Neck											х	
22	<u>2013262</u>	Consultation, Neuropathology											х	
23	<u>2013263</u>	Consultation, Surgical Pathology											х	
23	0060360	Corynebacterium diphtheriae Culture				x								
23	0020414	Creatine Kinase Isoenzymes					x							
24	2001613	Crohn Disease Prognostic Panel						х			х			
48	0055283	Cysticercosis Antibody, IgG by Western Blot												x



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48	<u>0055282</u>	Cysticercosis Antibody, IgG by Western Blot (CSF)												X
48	<u>2008920</u>	Cytochrome P450 Pain Management Panel, CYP2D6, CYP2C9, CYP2C19 - Common Variants												x
48	<u>0091589</u>	Darvocet, Urine												х
24	<u>2013294</u>	Dengue Virus (1-4) Subtype by PCR											Х	
24	<u>2002247</u>	Disaccharidase, Tissue			х		х							
25	<u>2006621</u>	Drug Detection Panel, Umbilical Cord Tissue, Qualitative		x			x	x			x	x		
48	<u>0051464</u>	Dysautonomia, Familial (<i>IKBKAP</i>) 2 Mutations, Fetal												x
27	<u>2008603</u>	<i>ERBB2</i> (<i>HER2</i> /neu) Gene Amplification by FISH, Tissue				x								
27	<u>2013277</u>	Esterase, Non-Specific Cytochemical Stain Only											Х	
27	<u>2012695</u>	Ethyl Glucuronide Screen Only, Urine					х							
48	<u>0051469</u>	Fanconi Anemia, Group C (FANCC) 2 Mutations, Fetal												x
48	<u>0030140</u>	Fibrin/Fibrinogen Degradation Split Products												х
48	<u>0091509</u>	Formic Acid, Urine												х
48	<u>0051439</u>	Gaucher Disease (GBA) 8 Mutations, Fetal												х
48	<u>0091353</u>	Glyburide Quantitative, Serum or Plasma												х
28	<u>0040080</u>	Hematocrit				х	х							
28	<u>0040085</u>	Hemoglobin				х	х							
28	<u>0020416</u>	Hepatic Function Panel										х		
28 29	<u>0092522</u> <u>2013333</u>	Histoplasma Antigen by EIA, Serum Human Immunodeficiency Virus (HIV) Combo Antigen/Antibody (HIV-1/O/2) by ELISA, with Reflex to HIV-1/HIV-2 Antibody Differentiation, Supplemental				X							x	
29	<u>2002899</u>	Human Papillomavirus (HPV), High Risk by in situ Hybridization, Paraffin									х			
30	<u>0050980</u>	Humoral Immunity Panel I										x		
30	<u>0050981</u>	Humoral Immunity Panel II										х		
31	<u>2013270</u>	Inflammatory Bowel Disease Differentiation Panel											х	
48	0050567	Inflammatory Bowel Disease Differentiation Profile												x
48	<u>0091530</u>	Inhalants Panel, Solvents, Serum or Plasma												x
31	<u>0049110</u>	Iron Stain				х								
31	0080200	Lecithin-Sphingomyelin Ratio				х								



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31	<u>2005661</u>	Liver Fibrosis, Chronic Viral Hepatitis (Echosens FibroMeter)										x		
32	<u>0040005</u>	Manual Differential					х							
33	<u>0081293</u>	Maternal Screening, Sequential, Specimen #1				х								
33	0081150	Maternal Serum Screen, First Trimester				х								
33	<u>0081062</u>	Maternal Serum Screening, Integrated, Specimen #1				х								
33	<u>2013305</u>	Meningitis/Encephalitis Panel by PCR											х	
34	2011521	Meprobamate, Serum or Plasma, Quantitative					х					х		
48	<u>0051449</u>	Mucolipidosis, Type IV (<i>MCOLN1</i>) 2 Mutations, Fetal												x
34	<u>2013273</u>	Myeloperoxidase, Cytochemical Stain Only											х	
48	2012535	Nerve Fiber Density Analysis, Intraepidermal												х
48	0051459	Niemann-Pick, Type A (<i>SMPD1</i>) 4 Mutations, Fetal												х
48	2008868	Nonalcoholic steatohepatitis (NASH) FibroSURE												х
34	2008767	Opioid Receptor, mu OPRM1 Genotype, 1 Variant		х	х	х		х						
35	<u>2007479</u>	Pain Management Drug Panel by High-Resolution Time-of-Flight Mass Spectrometry and Enzyme Immunoassay, Urine									x			
35	<u>2009288</u>	Pain Management Drug Screen with Interpretation by High-Resolution Time-of-Flight Mass Spectrometry and Enzyme Immunoassay, Urine									x			
35	<u>2012603</u>	PAX8-PPARG Translocations Detection by PCR			х									
35	<u>0091260</u>	Phenol Exposure Quantitative, Urine			х									
35	<u>2010481</u>	Phenytoin, Free					х							
35	<u>0090141</u>	Phenytoin, Free and Total					х							
35	<u>0040235</u>	Platelets					х							
35	<u>2003040</u>	PM/Scl-100 Antibody, IgG by Immunoblot with Reflex to ANA IFA (Pricing Change Only)												
35	<u>2002871</u>	<i>PML-RARA</i> Translocation, t(15;17) by RT-PCR, Quantitative						x						
36	<u>0095044</u>	Prenatal Reflexive Panel		х										
36	<u>2008509</u>	Progesterone Quantitative by HPLC-MS/MS, Serum or Plasma					x							
37	<u>2013352</u>	Pyridoxine-Dependent Epilepsy Panel, Serum or Plasma											x	
38	<u>2013355</u>	Pyridoxine-Dependent Epilepsy Panel, Urine											x	
38	0040270	Red Blood Cell Count				x	x							
39	0040263	Reticulocyte, Hemoglobin Panel	x			x	x				x			



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40	<u>0040022</u>	Reticulocytes, Percent and Number	х			х	х							
40	<u>2013358</u>	S100B, CSF											Х	
48	<u>2006462</u>	Scleroderma Antibodies Panel												х
41	<u>2013251</u>	STAT6 by Immunohistochemistry											х	
42	<u>0050725</u>	Streptococcus pneumoniae Antibodies, IgG (14 Serotypes)						x	x			x		
43	<u>2005779</u>	<i>Streptococcus pneumoniae</i> Antibodies, IgG (23 Serotypes)					x	x	x			x		
44	<u>2008919</u>	Streptococcus pneumoniae Antibodies, IgG (9 Serotypes)					x	x	x			x		
45	<u>2013325</u>	Systemic Scleroderma Comprehensive Panel											х	
46	<u>2013275</u>	Tartrate-Resistant Acid Phosphatase, Cytochemical Stain Only											x	
48	0051429	Tay-Sachs Disease (HEXA) 7 Mutations, Fetal												х
46	<u>2013335</u>	Thrombopoietin (TPO), Serum											х	
47	<u>2013290</u>	Tropheryma whipplei PCR											х	
48	<u>2011025</u>	Tropheryma whipplei Detection by PCR, Blood												x
48	<u>2008116</u>	Urine Culture, Invasive Collection												x
47	2005413	Urticaria-Inducing Activity				х								
48	<u>2004175</u>	Vascular Endothelial Growth Factor C (VEGF-C) by Immunohistochemistry												x
47	0080380	Vitamin C (Ascorbic Acid), Plasma				x								
47	0040320	White Blood Cell Count				х	х							
48	2011127	Zolpidem and Metabolites Quantitative, Urine												x



0060152 Acid-Fast Bacillus (AFB) Culture and AFB Stain

Specimen Required: Remarks: Specimen source required.

<u>Unacceptable Conditions:</u> Dry material or material collected and transported on a swab. Acid Fast Stain: Stool, blood, bone marrow, and grossly bloody specimens, CSF if less than 5 mL, or urine specimens if less than 40 mL.

Note: Positive cultures are reported as soon as detected. AFB stain, AFB identification of positives, and susceptibility tests are billed separately from culture. Identification of positive culture is billed by individual DNA probes and tests performed. *Mycobacterium tuberculosis* Complex Detection and Rifampin Resistance by PCR (ARUP test code 2010775) is available for respiratory, CSF or body fluid specimens.

The laboratory should be notified when the presence of *Mycobacterium genavense* is suspected as this organism will not grow on media routinely used for *Mycobacterium* isolation.

Susceptibility will be performed on organisms isolated from a sterile source and isolates of *Mycobacterium chelonae*, *M. abscesses*, *M. fortuitum complex*, *M. immunogenum*, *M. mucogenicum*. Susceptibility testing will be performed by request only on *M. kansaii* and *M. marinum*. Susceptibility testing of *M. gordonae* is inappropriate.

For AFB susceptibility information, refer to Antimicrobial Susceptibility - AFB Mycobacteria (ARUP test code 0060217).

For AFB culture on blood and bone marrow refer to Culture, Acid-Fast Bacillus, Blood (ARUP test code 0060060).

0020008Alanine Aminotransferase, Serum or PlasmaALT

HOT LINE NOTE: There is a unit of measure change associated with this test. Change unit of measure for component 0020008, Alanine Aminotransferase from IU/L to U/L.

<u>2005736</u>	Alkaline Phosphatase Isoenzymes, CSF	CSF ALKP I
Performed: Reported:	Sun-Sat 1-3 days	
0021020	Alkaline Phosphatase Isoenzymes, Serum or Plasma	ALKP-ISO

Specimen Required: Stability (collection to initiation of testing): After separation from cells: Ambient: 1 week; Refrigerated: 1 week; Frozen: 2 months

MC AFB



0060217 Antimicrobial Susceptibility, AFB/Mycobacteria

MA AFB

Reference Interval:

Available Separately	Test Name	Methodology	Reference Interval/Drugs Tested	CPT Code
Yes (0060347)	Antimicrobial Susceptibility - AFB/Mycobacterium tuberculosis Primary Panel	MGIT960	The interpretation provided is based on results for the following drugs at the stated concentrations: Drugs tested: Ethambutol: 5.0 $\mu g/mL$; Isoniazid: 0.1 $\mu g/mL$ (0.4 $\mu g/mL$ if resistant to 0.1 $\mu g/mL$; Rifampin: 1.0 $\mu g/mL$. This procedure screens isolates of <i>M. tuberculosis</i> complex for drug resistance. The procedure does not use serial dilutions to provide quantitative MIC	87188 x4
			values. Single critical concentrations for each antimycobacterial agent used have been defined by the United States Public Health Service.	
No	Antimicrobial Susceptibility - AFB/Mycobacterium tuberculosis Secondary Panel	Agar proportion and Broth dilution	Effective February 21, 2012 Note: If <i>M. tuberculosis</i> isolate is resistant to rifampin or any two primary drugs, a secondary panel will be performed as a send-out test. The interpretation provided is based on testing for the following drugs at the stated concentrations: Drugs tested: Amikacin: 6 $\mu g'mL$; capreomycin: 10 $\mu g'mL$; cycloserine: 60 $\mu g/mL$; ethionamide: 10 $\mu g'mL$; kanamycin: 6 $\mu g'mL$; PAS: 8 $\mu g'mL$; streptomycin at a low level (2.0 $\mu g/mL$) and a high level (4.0 $\mu g/mL$).	87190 x6, 87188 x3
No	Antimicrobial Susceptibility - AFB/Mycobacteria	Broth Microdilution	See organism-specific panels below.	87186



No	Mycobacterium avium-	Broth Microdilution	Effective May 16, 2016	87186
	intracellularae Complex			
			Drugs tested: Amikacin,	
			ciprofloxacin, clarithromycin,	
			doxycycline, ethambutol,	
			ethionamide, isoniazide,	
			linezolid, moxifloxacin,	
			rifabutin, rifampin	
			streptomycin and	
			(TMD/SVT)	
			(1MP/SA1).	
			Selective reporting by	
			organism.	
			Clarithromycin, moxifloxacin	
			and linezolid are the only drugs	
			for which CLSI provides	
			interpretive guidelines.	
			Clarithromycin results predict	
			azithromycin. For Amikacin	
			and Ciprofloxacin only MIC is	
			reported. Because MIC results	
			do not predict clinical response	
			and may be misleading,	
			athembutal MICs are not	
			routinely reported and must be	
			specifically requested	
No	Rapid Growing Mycobacteria	Broth Microdilution	Effective August 17, 2015	87186
			Drugs tested: Amikacin,	
			celoxitin, ciprofloxacin,	
			iminonam linegolid	
			minocycline moviflovacin	
			tigecycline tobramycin (M	
			chelonae only) and	
			trimethoprim/sulfamethoxazole	
			(TMP/SXT). Selective	
			reporting by organism.	



No	Other Slowly-Growing	Broth Microdilution	Effective May 20, 2013	87186
	Nontuberculosus Mycobacteria			
	(NTM)		Drugs tested: Amikacin,	
			ciprofiloxacin, clarithromycin,	
			doxycycline, etnambutol,	
			linezolid moxifloxacin	
			rifabutin rifampin	
			streptomycin and	
			trimethoprim/sulfamethoxazole	
			(TMP/SXT). Selective	
			reporting by organism.	
			CLSI recommends that isolates	
			of <i>M. kansasu</i> be tested against	
			only Rifampin-susceptible	
			isolates are also susceptible to	
			rifabutin. If the isolate is	
			rifampin-resistant, the	
			following secondary drugs will	
			also be reported: Amikacin,	
			linezolid moviflovacin	
			rifabutin streptomycin and	
			trimethoprim-	
			sulfamethoxazole.	
			M. marinum isolates are tested	
			against amikacin, ciprofloxacin,	
			clarithromycin, doxycycline,	
			rifebutin rifempin and	
			trimethoprim-	
			sulfamethoxazole.	
			Interpretation is based on CLSI	
			guidelines.	
			Slowly-growing NTM other	
			than M. kansasii and M.	
			marinum are tested against	
			amikacin, ciprofioxacin,	
			linezolid moxifloxacin	
			rifabutin, rifampin,	
			streptomycin, and	
			trimethoprim-	
			sulfamethoxazole.	
			Interpretive criteria are based	
			on CLSI guidelines for M. kansasii.	



New Test	<u>2013341</u>	Apolipoprotein E (APOE) Genotyping, Alzheimer Disease Risk	APOE AZ
Available July 5	5, 2016		

Methodology:Polymerase Chain Reaction/Fluorescence MonitoringPerformed:Mon, ThuReported:2-7 days

Specimen Required: Patient Prep:

Collect: Lavender (EDTA), Pink (K₂EDTA), or Yellow (ACD Solution A or B). <u>Specimen Preparation:</u> Transport 3 mL whole blood. (Min: 1 mL) <u>Storage/Transport Temperature:</u> Refrigerated. <u>Remarks:</u> Testing of fetal specimens or specimens from patients under the age of 18 years is not offered. <u>Unacceptable Conditions:</u> Plasma or serum. Heparinized specimens. <u>Stability (collection to initiation of testing):</u> Ambient: 72 hours; Refrigerated: 2 weeks; Frozen: 1 month.

Reference Interval: Homozygous apo e3 (e3/e3): This genotype is the most common (normal) genotype.

Interpretive Data:

Background Information for Apolipoprotein E (APOE) Genotyping, Alzheimer Disease Risk

Characteristics: Alzheimer disease (AD), the most common cause of dementia, is characterized by progressive cognitive decline including memory, problem-solving skills, multi-step tasks, planning, and changes in personality. A clinical diagnosis of probable AD can be made based on clinical signs and neuroimaging, and the diagnosis is confirmed postmortem based on neuropathologic findings. The e4 allele of the *APOE* gene has been widely demonstrated to be associated with increased risk of AD. In individuals with a clinical diagnosis of AD, the presence of the e4 allele increases the likelihood that the diagnosis is correct, but is not diagnostic alone. *APOE* genotyping is not recommended for predicting AD risk in asymptomatic individuals. **Prevalence of** *APOE* e4: Heterozygosity and homozygosity for the e4 allele is present in approximately 25 percent and 1-2 percent of the general population, respectively.

Inheritance of APOE e4: Semi-dominant.

Penetrance of *APOE* **e4**: Incomplete and influenced by age, gender, ethnicity, family history and environmental factors. The e4 allele is neither necessary nor sufficient for diagnosing AD; therefore, not all individuals with AD have the e4 allele and not all individuals with the e4 allele will develop AD. **Cause:** Multi-factorial.

Variants Tested: Two single nucleotide polymorphisms in the *APOE* gene at codons 130 (rs429358) and 176 (rs7412). The e3 allele (Cysteine at 130 and Arginine at 176) is the most common in the general population. The e4 allele (Arginine at 130 and 176) is associated with increased AD risk. The e2 allele (Cysteine at codons 130 and 176) may be associated with a lower risk for AD but homozygosity has been associated with increased risk for type III hyperlipoproteinemia.

Clinical Sensitivity: Approximately 30-60 percent of individuals diagnosed with AD carry at least one e4 allele. The e4/e4 genotype is found in approximately 13 percent of the AD population and 20 percent of the familial AD population.

Methodology: Polymerase chain reaction (PCR) and fluorescence monitoring using hybridization probes.

Analytical Sensitivity and Specificity: 99 percent.

Limitations: Only the APOE alleles e2, e3 and e4 will be detected; rare alleles are not detected by this test. Diagnostic errors can occur due to rare sequence variations.

This test is performed pursuant to an agreement with Roche Molecular Systems, Inc. See Compliance Statement C: www.aruplab.com/CS

CPT Code(s): 81401

New York DOH approval pending. Call for status update.



New Test	<u>2013337</u>	Apolipoprotein E (APOE) Genotyping, Cardiovascular Risk	APOE CR
Available July 5,	2016		
Methodology: Performed: Reported:	Polymerase Chai Mon, Thu 2-7 days	n Reaction/Fluorescence Monitoring	
Specimen Required	Example 2 Patient Prep: <u>Collect:</u> Lavende <u>Specimen Prepar</u> <u>Storage/Transpor</u> <u>Remarks:</u> This te <u>Unacceptable Co</u> <u>Stability (collect</u>	er (EDTA), Pink (K ₂ EDTA), or Yellow (ACD Solution A or B). <u>ation:</u> Transport 3 mL whole blood. (Min: 1 mL) <u>rt Temperature:</u> Refrigerated. st is not recommended for nonsymptomatic patients under 18 years of age. <u>inditions:</u> Plasma or serum. Heparinized specimens. <u>ion to initiation of testing):</u> Ambient: 72 hours; Refrigerated: 2 weeks; Frozen: 1 month	
Reference Interva	l: Homozygous A	POE e3 (e3/e3): This genotype is the most common (normal) genotype.	
Interpretive Data Background Inform Characteristics: Hyp premature vascular di Incidence of HPL II Inheritance of HPL II Inheritance of HPL II Inheritance of HPL II Inheritance of HPL II Operation of HPL II Inheritance of HPL II Inheritance of HPL II Inheritance of HPL II Cause: 2 copies of th necessary nor sufficie Variants Tested: AF p.Cys130Arg and c.5 Clinical Sensitivity: Methodology: Polyn Analytical Sensitivit Limitations: Only th isoelectric focusing n	ation for Apolipo perlipoproteinemia isease including cc I: Approximately III: Multifactoria ulso play a large ro ercent of individua e e2 allele provide ent for HPL III. OE gene alleles, e 26C, p.176Arg). 90 percent of indi perase chain reacti y and Specificity e e2, e3 and e4 va hay be indicated. I	 pprotein E (<i>APOE</i>) Genotyping, Cardiovascular Risk III (HPL III) is characterized by increased cholesterol and triglyceride levels, presence of B oronary heart disease (CHD) and peripheral artery disease. 1 in 5,000. I; greater than 90 percent of affected individuals are homozygous for the e2 allele but other fole in development of disease. Ils homozygous for the e2 will develop HPL III. es supporting evidence for a diagnosis of HPL III in a symptomatic individual but e2 homozy 2 (c.388T, p.130Cys and c.526C>T, p.Arg176Cys), e3 (c.388T, p.130Cys and c.526C, p.176 viduals with HPL III are homozygous for the e2 variant. on (PCR) and fluorescence monitoring using hybridization probes. : 99 percent. riants will be detected. Rare isoforms of <i>APOE</i> will not be detected. If rare alleles are suspecting of the rare sequence variations. 	-VLDL, xanthomas, and actors such as diabetes ygosity is neither 5Arg), e4 (c.388T>C, cted, phenotyping by
See Compliance State	ement C: www.art	plab.com/CS	
CPT Code(s):	81401		
New York DOH App	roved.		
HOT LINE NOTI	E: Refer to the Te	st Mix Addendum for interface build information.	



New Test2013327Aquaporin-4 Receptor Antibody by ELISA with Reflex toAQEAquaporin-4 Receptor Antibody, IgG by IFA

Available April 18, 2016

Methodology:Semi-Quantitative Enzyme-Linked Immunosorbent Assay/ Semi-Quantitative Indirect Fluorescent AntibodyPerformed:Tue, ThuReported:1-6 days

Specimen Required: Collect: Serum Separator Tube (SST).

 Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum to an ARUP Standard Transport Tube. (Min: 0.3 mL)

 Storage/Transport Temperature: Refrigerated.

 Unacceptable Conditions: Amniotic fluid, CSF, pericardial fluid, ocular fluid, peritoneal fluid, synovial fluid, or plasma.

 Contaminated, hemolyzed, icteric, or lipemic specimens.

 Stability (collection to initiation of testing):

 After separation from cells: Ambient: 72 hours; Refrigerated: 2 weeks; Frozen: 1 month (avoid repeated freeze/thaw cycles)

Reference Interval:

Components	Reference Interval
Aquaporin-4 Receptor Antibody	Negative: 4 U/mL or less
	Indeterminate: 5 U/mL
	Positive: 6 U/mL or greater
Aaquaporin-4 Receptor Antibody, IgG by IFA, Serum with Reflex to Titer	Less than 1:10

Interpretive Data: Approximately 75 percent of patients with neuromyelitis optica (NMO) express antibodies to the aquaporin-4 (AQP4) receptor. Diagnosis of NMO requires the presence of longitudinally extensive acute myelitis (lesions extending over 3 or more vertebral segments) and optic neuritis. While absence of antibodies to the AQP4 receptor does not rule out the diagnosis of NMO, presence of this antibody is diagnostic for NMO.

See Compliance Statement D: www.aruplab.com/CS

NOTE: If AQP4 antibody IgG by ELISA is positive, then AQP4 antibody IgG by IFA will be added. If AQP4 antibody IgG by IFA is positive, then an AQP4 antibody IgG titer will be added. Additional charges apply.

CPT Code(s): 83516; if reflexed, add 86255; if reflexed, add 86256

New York DOH approval pending. Call for status update.

HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

AQP4 R



New Test2013320Aquaporin-4 Receptor Antibody, IgG by IFA with Reflex to Titer,AQP4 SERSerum

Available April 18, 2016

Methodology:	Semi-Quantitative Indirect Fluorescent Antibody
Performed:	Wed
Reported:	1-8 days

 Specimen Required:
 Collect:
 Serum Separator Tube (SST) or Plain Red.

 Specimen Preparation:
 Transfer 1 mL serum to an ARUP Standard Transport Tube. (Min: 0.15 mL)

 Storage/Transport Temperature:
 Refrigerated.

 Unacceptable Conditions:
 Contaminated.

 Stability (collection to initiation of testing):
 Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

Reference Interval: Less than 1:10

Interpretive Data: Diagnosis of neuromyelitis optica (NMO) requires the presence of longitudinally extensive acute myelitis (lesions extending over 3 or more vertebral segments) and optic neuritis. Approximately 75 percent of patients with NMO express antibodies to the aquaporin-4 (AQP4) receptor. While the absence of AQP4 receptor antibodies does not rule out a diagnosis of NMO, presence of this antibody is diagnostic for NMO.

See Compliance Statement D: www.aruplab.com/CS

Note: If AQP4 antibody IgG is positive, then an AQP4 antibody IgG titer is reported. Additional charges apply.

CPT Code(s): 86255; if reflexed, add 86256

New York DOH approval pending. Call for status update.

HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

0095227Arylsulfatase A, 24-Hour UrineARYLSULF AMethodology:Quantitative Colorimetry/Enzyme ImmunoassayPerformed:VariesReported:9-17 daysHOT LINE NOTE: There is a component change associated with this test.

Add component 2013301, Arylsulfatase A – Interpretation Add component 2013302, Arylsulfatase A - Reviewed By

0020007 Aspartate Aminotransferase, Serum or Plasma

HOT LINE NOTE: There is a unit of measure change associated with this test. Change unit of measure for component 0020007, Aspartate Aminotransferase from IU/L to U/L AST



2008901 B-Cell Memory and Naive Panel

CVID

Reference Interval: Effective May 16, 2016

Available Separately	Componen ts	Reference Interval							
		2-5 months	5-9 months	9-15 months	15-24 months	2-5 years	5-10 years	10-16 years	16 years and older
No	B-cells % CD19	18-38 %	16-34 %	14-28 %	16-34 %	14-29 %	10-24 %	9-23 %	5-26 %
No	B-cells Absolute CD19	700-2400 cells/µL	700-2800 cells/µL	400-2900 cells/µL	600-1900 cells/μL	400-1700 cells/μL	300-600 cells/µL	200-600 cells/µL	58-558 cells/µL
No	Naive B-cell % CD19+/CD27 -/IgD+	82-95 %	86-93 %	77-95 %	68-89 %	54-88 %	47-77 %	51-83 %	29-93 %
No	Naive B-cell Absolute CD19+/CD27 -/IgD+	620-2120 cells/μL	600-2590 cells/μL	360-2800 cells/µL	490-1560 cells/μL	280-1330 cells/µL	130-460 cells/µL	120-430 cells/µL	22-423 cells/µL
No	Class- switched Mem%CD19 +/CD27+/IgD -/IgM-	0-9 %	2-7 %	1-12 %	4-14 %	5-21 %	11-30 %	9-26 %	3-23 %
No	Class- switched Abs CD19+/CD27 +/IgD-/IgM-	10-170 cells/μL	20-140 cells/µL	10-100 cells/µL	30-180 cells/µL	20-220 cells/µL	40-140 cells/µL	30-110 cells/µL	4-62 cells/µL
No	Non- Switched Mem %CD19+/CD 27+/IgD+/Ig M+	3-9 %	3-7 %	3-11 %	4-14 %	3-20 %	5-20 %	5-18 %	2-25 %
No	Non-switched Abs CD19+/CD27 +/IgD+/IgM+	20-200 cells/µL	30-120 cells/µL	20-140 cells/µL	30-170 cells/µL	20-180 cells/µL	20-100 cells/µL	20-70 cells/µL	4-66 cells/µL
No	Total Memory B- cell % CD19+/CD27 +	3-12 %	5-12 %	4-21 %	10-27 %	8-37 %	19-47 %	13-48 %	7-48 %
No	Total Memory B- cell Absolute CD19+/CD27 +	40-230 cells/µL	50-270 cells/μL	40-190 cells/µL	50-330 cells/µL	50-390 cells/µL	60-230 cells/μL	50-200 cells/µL	13-148 cells/μL
No	IgM Only Memory Pct CD19+CD27 +1gD-IgM+	0.3-6.0 %	0.3-6.0 %	0.3-6.0 %	0.3-6.0 %	0.3-6.0 %	0.3-6.0 %	0.3-6.0 %	0.3-6.0 %
No	IgM Only Memory Abs CD19+CD27 +IgD-IgM+	0.6-16.4 cells/μL	0.6-16.4 cells/μL	0.6-16.4 cells/µL	0.6-16.4 cells/µL	0.6-16.4 cells/µL	0.6-16.4 cells/µL	0.6-16.4 cells/µL	0.6-16.4 cells/μL

Interpretive Data: This panel is indicated for patients with suspected immune deficiencies, especially Common Variable Immune Deficiency (CVID), and to assess reconstitution of B-cell subsets after bone marrow or stem cell transplant. Subsets measured: B-cells (CD19+), total memory B-cells (CD19+ CD27+), class switched memory B-cells (CD19+ CD27+ IgD- IgM-), non-switched/marginal zone memory B-cells (CD19+ CD27+ IgD+ IgM+), IgM only memory B-cells (CD19+ CD27+ IgD-IgM+), and naive B-cells (CD19+ CD27-IgD+).

See Compliance Statement A: www.aruplab.com/CS

Note: Reference intervals for IgM-only memory B-cells have currently been established only for populations aged 16-years and older. For all other B-cell subsets, reference intervals for populations younger than 16-years are adopted from literature. Piątosa B, Wolska-Kuśnierz B, Pac M, Siewiera K, Gałkowska E, Bernatowska E. B cell subsets in healthy children: Reference values for evaluation of B cell maturation process in peripheral blood. Cytometry Part B 2010; 78B: 372–381.

CPT Code(s): 86355; 86356 x3



HOT LINE NOTE: There is a component change and a clinically significant name change associated with this test.

Change the charting name of component 2008904 from Class-switched Memory %CD19+/CD27+/IgD- to Class-switched Memory %CD19+/CD27+/IgD-/IgM-

Change the charting name of component 2008905 from Class-switched Absolute CD19+/CD27+/IgD- to Class-switched Absolute CD19+/CD27+/IgD-/IgM-

Change the charting name of component 2008906 from Non-Switched Memory %CD19+/CD27+/IgD+ to Non-Switched Memory %CD19+/CD27+/IgD+/IgM+

Change the charting name of component 2008907 from Non-switched Absolute CD19+/CD27+/IgD+ to Non-switched Absolute CD19+/CD27+/IgD+/IgM+ Add component 2013303, IgM Only Memory Abs CD19+CD27+IgD-IgM+

Add component 2013304, IgM Only Memory Pct CD19+CD27+IgD-IgM+

BCRABL NGS 2008420 BCR-ABL1 Mutation Analysis for Tyrosine Kinase Inhibitor Resistance by Next **Generation Sequencing**

Note: For specimens having no t(9;22) fusion, this test will be canceled and the ABL1 Amplification Confirmation test will be ordered.

2005017	BCR-ABL1, Major (p210), Quantitative	BCR MAJ
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HOT LINE NOTE: There is a component change associated with this test.

Remove component 2005271, BCR-ABL1 Maj Molecular Response achieved

2005010 BCR-ABL1, Qualitative with Reflex to BCR-ABL1 Quantitative

HOT LINE NOTE: There is a component change associated with this test.

Reflex component 2005011, BCR-ABL1, Major, Quant has a component change. Remove component 2005271, BCR-ABL1 Maj Molecular Response achieved.

2011450 Carisoprodol and Meprobamate, Serum or Plasma, Quantitative

Reference Interval:

Test Number	Components	Reference Interval	
	Carisoprodol, Serum or	Less than 8.0 µg/mL	
	Plasma	Toxic: Greater than or equal to 8.0 µg/mL	
	Meprobamate, Serum or		
	Plasma		
		Dose-Related Range	5.0-20.0 µg/mL
		Toxic	Greater than 40.0 µg/mL

HOT LINE NOTE: There is a numeric map change associated with this test that affects interface clients only.

Change the numeric map for component 2011455 from XXXX to XXXX.X Change the numeric map for component 2011456 from XXXX to XXXX.X BCR RFLX

CARIS SP



0040002 **CBC** with Platelet Count

CBC

Specimen Required: Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 48 hours; Frozen: Unacceptable

Kultitut	e miter var. Ener	11ve wiay 10, 20	510											
Test Number	Components	Reference Inter	val											
0040080	Hematocrit	Age	0	1-6	7-13	14-29	30-60	61-90	91-180	6-23	2-5	6-11	12-17	18 years
		Ŭ	days	days	days	days	days	days	days	months	years	years	years	and older
		Male %	42-60	45-64.9	42-64.9	39-63	31-55	28-42	29-41	33-39	34-40	35-45	37-49	44.2-53
		Female %	42-60	45-64.9	42-64.9	39-63	31-55	28-42	29-41	33-39	34-40	35-45	36-46	36-49
0040085	Hemoglobin	Age	0	1-6	7-13	14-29	30-60	61-90	91-180	6-23	2-5	6-11	12-17	18 years
	J	0	days	days	days	days	days	days	days	months	years	years	years	and older
		Male (g/dL)	13.5-19.5	14.5-22.5	13.5-21.5	12.5-20.5	10.0-18.0	9.0-14.0	9.5-13.5	10.5-13.5	11.5-13.5	11.5-15.5	13.0-16.0	14.8-17.8
		Female (g/dL)	13.5-19.5	14.5-22.5	13.5-21.5	12.5-20.5	10.0-18.0	9.0-14.0	9.5-13.5	10.5-13.5	11.5-13.5	11.5-15.5	12.0-16.0	12.6-15.9
0040270	Red Blood Cell	Age	0	1-6	7-13	14-29	30-60	61-90	91-180	6-23	2-5	6-11	12-17	18 years
	Count	0	days	days	days	days	days	days	days	months	years	years	years	and older
		Male (M/µL)	3.9-5.5	4.0-6.6	3.9-6.3	3.6-6.2	3.0-5.4	2.7-4.9	3.1-4.5	3.7-5.3	3.9-5.3	4.0-5.2	4.5-5.3	4.7-6.14
		Female (M/µL)	3.9-5.5	4.0-6.6	3.9-6.3	3.6-6.2	3.0-5.4	2.7-4.9	3.1-4.5	3.7-5.3	3.9-5.3	4.0-5.2	4.1-5.1	4.08-5.47
0040320	White Blood Cell	Age	0	1-6	7-13	14-29	30-60	2-11	1-3	4-5	6-7	8-13	14-17	18 years
	Count	9.	days	days	days	days	days	months	years	years	years	years	years	and older
		Male (K/µL)	9-30	9-34	5-21	5-20	5-19.5	5.5-17	6-17	5.5-15.5	5-14.5	4.5-13.5	4.5-13	4.3-11.3
		Female (K/µL)	9-30	9-34	5-21	5-20	5-19.5	5.5-17	6-17	5.5-15.5	5-14.5	4.5-13.5	4.5-13	4.3-11.3
	RDW	11.5-15.3 %												
	MPV	8.6-12.3 fL												
	MCV	Age	0	1-6	7-13	14-29	30-60	61-90	91-180	6-23	2-5	6-11	12-17	18 years
		0	days	days	days	days	days	days	days	months	years	years	years	and older
		Male (fL)	98-118	95-121	88-126	86-124	85-123	77-115	74-108	70-86	75-87	77-95	78-98	81.2-96.6
		Female (fL)	98-118	95-121	88-126	86-124	85-123	77-115	74-108	70-86	75-87	77-95	78-102	81.9-101
	MCH	Age	0-6	7-29	30-60	61-180	6-23	2-5	6-11	12-17	18 years a	nd older	-	
		Ŭ	days	days	days	days	months	years	years	years				
		Male (pg)	31-37	28-40	26-34	25-35	23-31	24-30	25-33	25-35	25.8-33.1			
		Female (pg)	31-37	28-40	26-34	25-35	23-31	24-30	25-33	25-35	25.8-33.1			
	MCHC	Male (g/dL)	31.9-35.2											
		Female (g/dL)	31.2-34.5											
	NRBC	Age	0-3 days	4 days and	older									
		% (/100 WBC)	0.1-8.3	0										
		K/µL	0-1.3	0										
0040235	Platelets	159-439 K/µL												
	IPF	Age	0-180	6-23	2-5	6-11	12-17	18 years	and older					
		8	days	months	years	years	years							
		Male (%)	2.3-7.1	1.7-4.1	1.4-3.9	1.3-5.2	1.9-6.4	1-11.4						
		Female (%)	1.6-7.1	1.7-4.8	1.3-3.9	1.3-5.0	1.7-6.7	1-11.4						
				-	-		-							

Reference Interval: Effective May 16, 2016

HOT LINE NOTE: There is a component change associated with this test.

Add component 2013370, Nucleated Red Blood Cells % Add component 2013369, Nucleated Red Blood Cells #

Add component 2013361, Immature Platelet Fraction



0040003 CBC with Platelet Count and Automated Differential

CBCAD

Specimen Required: Stability (collection to initiation of testing): Ambient: 24 hours (without smears); Refrigerated: 48 hours; Frozen: Unacceptable

`est **Reference Interval** Components Number 0040080 14-29 91-180 Hematocrit Age **1-6** 7-13 80-60 51-90 5-23 2-5 5-11 12-17 18 years and older days nonth days lays ear ear ears 33-39 4.2-53 Male % 12-60 45-64.9 42-64.9 9-63 31-55 28-42 29-41 84-40 35-45 7-49 Female % 12-60 45-64.9 12-64.98-42 9-41 3-39 34-40 35-45 6-46 36-49 0040085 Hemoglobin Age 1-6 7-13 14-29 30-60 61-90 91-180 6-23 2-5 6-11 12-17 18 years 0 iontl nd older ays lays Male (g/dL) 13.5-19. 13.5-21. 10.0-18.0 10.5-13.5 13.0-16.0 14.5-22.5 12.5-20. 9.0-9.5-13.5 11.5-13.5 11.5-15.5 14.8-17.8 14.0 Female (g/dL) 13.5-19.5 14.5-22.5 13.5-21.5 12.5-20.5 10.0-18.0 9.0-9.5-13.5 10.5-13.5 11.5-13.5 11.5-15.5 12.0-16.0 12.6-15.9 400040270 Red Blood Cell 14-29 61-90 \ge 1-6 7-13 30-60 91-180 5-23 2-5 5-11 12-17 l8 vears and older nonths Count days avs lavs ears ear ears lavs avs Male (M/µL) .9-5.5 4.0-6.6 3.9-6.3 3.6-6.2 3.0-5.4 7-4.9 3.1-4.5 .7-5.3 3.9-5.3 4.0-5.2 .5-5.3 4.7-6.14 Female (M/µL) .9-5 4.0-6.6 3.9-6 8.6-6 3.0-5.4 .7-4.9 3.1-4.5 3.9-5 4.0-5. 4.08-5.47 3.7-5.3 4.1-5. 0040320 White Blood Cell 7-13 14-29 30-60 2-11 1-3 14-17 1-6 4-5 5-7 8-13 18 years Age Count days days nonth and older lays lays lays years years ears years ears Male (K/µL) -30 9-34 -21 -20 -19.5 .5-17 5-17 .5-15.5 5-14.5 4.5-13. .5-13 4.3-11.3 9-34 5-21 -20 Female (K/uL) 9-30 5-19 5 5-17 5-17 5 5-15 5 5-14 5 4 5-13 5 5-13 4 3-11 3 2DW 11 5-15 3 % .6-12.3 fL MPV 7-13 MCV 1-6 14-29 30-60 61-90 91-180 6-23 2-5 6-11 12-17 18 years Age 0 days nonth and older days lay day day lay ears ears 1.2-96.6 Male (fL) 95-12 4-10 0-80 8-11 5-12 8-12 6-12 7-115 emale (fl 8-11 95-12 6-12 5-10 15-87 81 9-101 8-12 7-114 4-10 70-86 8-10 MCH 7-29 30-60 12-17 0-6 61-180 5-23 2-5 5-11 18 years and older Age nonth lay lays lays lays ears ear /ears Male (pg) 28-40 6-34 3-31 4-30 5-33 5-35 25 8-33 1 28-40 26-34 5-35 23-31 24-30 5-33 25-35 25.8-33.1 Female (pg) 31.9-35 MCHC Male (g/dL) Female (g/dL) 31.2-34.5 NRBC 0-3 days 4 days and older Age % (/100 WBC 0.1-8 K/µL 0 - 1.30-11 1-5 6-13 14-17 18 years and older Granulocytes Age 1 Number month vear ears K/μL .5-10.0 1.5-8.5 1.5-8.0 1.8-8.0 2.0-7.40-13 14-29 30-90 91-180 6-9 10-11 12-23 2-3 4-5 6-7 8-9 10-11 ١ge Granulocytes Percentage nonths nonths lays lays lays nonth ears /ears /ears vears ears lays 19-49 14-44 15-25 14-24 13-23 12-22 13-33 15-35 23-45 32-54 34-56 31-61 12-13 14-17 18 years and older ١ge ears ears Eosinophils ١ge 0-6 7 davs -11 1-13 14 vears and older Number nonths lays ears ζ/uL 0 - 10.1-1.1 0-07 0-0 5 Eosinophils 0.4-6.7% ercent Basophil 0-0.1 K/µL Jumbe Basophil 0.3-1.4% Aonocytes 0-6 days 1-5 6-17 18 years and older ١ge 11 months Number dav vears ears 0-0.8 K/µL 0.3-2.7 0-1.1 0.3 - 1.00.4 - 3.60-6 7-29 **30-6**0 61-120 4 months and older Monocytes \ge Percentage day lays lays lavs 0-9 0-12)-10)-9 4.1-12.4 6 18 years and older 0-6 7 days -1-5 5-13 14-17 Lymphocytes Age 1 month Jumber ears 2-17.0 1.5-7.0 1.2-5.8 4-10.5 1.3-3.6 K/uL

Reference Interval: Effective May 16, 2016



	Lymphocytes Percentage	Age	0-6 days	7-13 days	14-29 days	30-60 days	61-90 days	91-180 days	6-7 months	8-9 months	10-11 months	12-23 months	2-3 vears	4-5 vears
	rereennage	%	26-36	36-46	43-53	41-71	42-72	44-74	46-76	47-77	48-78	46-76	44-74	35-65
		Age	6-7	8-9	10-13	14-15	16-17	18 years a	nd older					
		%	years 27-57	years 24-54	years 28-48	years 27-47	years 25-45	17.6-49.6						
	Immature Granulocyte	Age	0-2 days	3-90 days	91-180 days	6-23 months	2-5 years	6-11 years	12-17 years	18 years	and older			
	Number	K/µL	0.08-1.68	0.03-0.71	0.02-0.09	0.02-0.18	0.02-0.09	0.02-0.06	0.02-0.05	0.01-0.09	9			
	Immature Granulocyte	Age	0-2 days	3-90 days	91-180 days	6-23 months	2-5 years	6-17 years	18 years a	nd older				
	percent	%	0.7-7.8	0.4-5.3	0.1-0.7	0.1-1.2	0.1-1.1	0.1-0.5	0.2-0.9					
0040235	Platelets	159-439 K/µL												
	IPF	Age	0-180 days	6-23 months	2-5 years	6-11 years	12-17 years	18 years a	nd older					
		Male (%)	2.3-7.1	1.7-4.1	1.4-3.9	1.3-5.2	1.9-6.4	1-11.4						
		Female (%)	1.6-7.1	1.7-4.8	1.3-3.9	1.3-5.0	1.7-6.7	1-11.4						

HOT LINE NOTE: There is a component change associated with this test.

Remove component 0040012, Lymphocyte # Remove component 0040013, Lymphocyte % Add component 2013370, Nucleated Red Blood Cells % Add component 2013369, Nucleated Red Blood Cells # Add component 2013434, Immature Granulocytes %

Add component 2013435, Immature Granulocyte #

Add component 2013361, Immature Platelet Fraction

Add component 0040125, Lymphocyte #

Add component 0040130, Lymphocyte %

2012717 CHARGE Syndrome (CHD7) Sequencing, Fetal

CHD7 FE

Specimen Require	 Collect: Cultured Amniocytes: Four T-25 flasks at 80 percent confluent cultured amniocytes. If the client is unable to culture amniocytes, this can be arranged by contacting Client Services at (800) 522-2787. Or Amniotic Fluid: Amniotic fluid submissions will require cultured amniocytes (Order Amniocyte Culture, Four Flask). AND Maternal Whole Blood: Lavender (K₂EDTA), Lavender (K₃EDTA), Pink (K₂EDTA), or Yellow (ACD Solution A or B). Specimen Preparation: Cultured Amniocytes: Fill flask with culture media. Transport four T-25 flasks at 80 percent confluent of culture amniocytes filled with culture media. Amniotic Fluid: Transport 10 mL unspun fluid (Min: 5 mL)
	Maternal Whole Blood: Transport 3 mL whole blood (Min: 1 mL)
0060850	Chlamydia trachomatis Culture CHLAM
Performed:	Sun-Sat
Reported:	2-5 days
Specimen Require	xd:Specimen Preparation: Immediately place swab, fluid, or washing in 3 mL universal transport medium such as M4, M4RT, M5, M6, UniTranz-RT, or UTM (ARUP supply #12884). Available online through eSupply using ARUP Connect [™] or contact ARUP Client Services et (800) 522 2787

Services at (800) 522-2787. <u>Unacceptable Conditions:</u> Urine. Specimens in any transport media other than indicated. Calcium alginate, dry, or wood swabs. <u>Stability (collection to initiation of testing)</u>: Ambient: 1 hour; Refrigerated: 48 hours; Frozen at -20°C: Unacceptable; Frozen at -70°C: 1 month

Note: Nucleic acid amplification testing is recommended for detection of *Chlamydia trachomatis* from endocervical or urethral specimens. Refer to *Chlamydia trachomatis* by Transcription-Medicated Amplification (TMA) (ARUP test code 0060243). Specimen must be collected and transported with test-specific kit.

0080469 Chromogranin A

CGA SERUM

Interpretive Data: This test is performed using the Cisbio CGA-ELISA-US kit. Results obtained with different methods or kits cannot be used interchangeably.

See Compliance Statement D: www.aruplab.com/CS



Clostridium difficile Cytotoxin Cell Assay	CDIFTX
Sun-Sat 2-5 days	
2013259 Comprehensive Kidney Biopsy Workup	KID REQ
ril 18, 2016	
Anatomic Pathology Test Request Form Recommended (ARUP form #32960) Time Sensitiv	e
Microscopy/Histochemistry/Immunofluorescence/Electron Microscopy	
Sun-Sat	
1-5 days	
red: <u>Collect</u> : Three renal needle core biopsies. <u>Specimen Preparation</u> : Obtain Renal Biopsy Collection Kit prior to collection procedure (AR through eSupply using ARUP Connect [™] or contact Client Services at (800) 522-2787. Speci instructions are provided with the kit. One biopsy placed in 10 percent formalin, one placed in Zeus fixative and one placed in gluta <u>Storage/Transport Temperature</u> : Room temperature. Also acceptable: Refrigerated. Ship in co <u>Remarks</u> : Submit electronic request. If you do not have electronic ordering capability, use an (#32960) with an ARUP client number. For additional technical details, contact ARUP Client clinical history. <u>Unacceptable Conditions</u> : Specimens submitted with non-representative tissue type. Renal tis <u>Stability (collection to initiation of testing)</u> : Ambient: 24 hours; Refrigerated: 24 hours; Froze	UP supply #40460) available online al fixatives are required; collection araldehyde. boled container during summer months. ARUP Anatomic Pathology Form t Services at (800) 522-2787. Submit ssue containing no glomeruli. en: Unacceptable
	Clostridium difficile Cytotoxin Cell Assay Sun-Sat 2-5 days 2013259 Comprehensive Kidney Biopsy Workup il 18, 2016 Anatomic Pathology Test Request Form Recommended (ARUP form #32960) Wicroscopy/Histochemistry/Immunofluorescence/Electron Microscopy Sun-Sat 1-5 days red: Collect: Three renal needle core biopsies. Specimen Preparation: Obtain Renal Biopsy Collection Kit prior to collection procedure (AR through eSupply using ARUP Connect™ or contact Client Services at (800) 522-2787. Speci instructions are provided with the kit. One biopsy placed in 10 percent formalin, one placed in Zeus fixative and one placed in gluta Storage/Transport Temperature: Room temperature. Also acceptable: Refrigerated. Ship in co Remarks: Submit electronic request. If you do not have electronic ordering capability, use an (#32960) with an ARUP client number. For additional technical details, contact ARUP Clien clinical history. Unacceptable Conditions: Specimens submitted with non-representative tissue type. Renal tis Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 24 hours; Froze

Interpretive Data: Refer to report.

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Note: Detailed collection instructions are available in the Renal Biopsy Collection Kit (#40460) or can be requested by contacting ARUP Client Services at (800) 522-2787. Use of a different collection kit could result in suboptimal biopsy fixation and delays in diagnosis. All vials must be labeled with the patient's full name and unique identifier. Avoid drying of specimens; make sure the specimens are completely submerged into the fixative and not caught on vial sides or in the threads of the cap.

NOTE: Ancillary testing is ordered at the discretion of the ARUP pathologist, additional CPT codes and charges may apply.

CPT Code(s): 88305, 88313 x3, 88346, 88350 x6

New York DOH Approved.

HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

0020408 Comprehensive Metabolic Panel

СМР

HOT LINE NOTE: There is a unit of measure change associated with this test. Change unit of measure for component 0020007, Aspartate Aminotransferase from IU/L to U/L Change unit of measure for component 0020008, Alanine Aminotransferase from IU/L to U/L



New Test Available Apri	2013260Comprehensive Muscle Biopsy Workup118, 2016	MUS REQ
	Anatomic Pathology Test Request Form Recommended (ARUP form #32960)	
Methodology: Performed: Reported:	Tissue Workup Sun-Sat 1-7 days	
Specimen Requir	ed: <u>Collect:</u> Muscle biopsy. <u>Specimen Preparation:</u> Obtain Muscle/Nerve Biopsy collection kit prior to collection procedure, (ARU online through eSupply using ARUP Connect [™] or contact Client Services at (800)522-2787. Special f instructions are provided with the kit. <u>Storage/Transport Temperature:</u> Room temperature. Also acceptable: Refrigerated. Ship in cooled cont <u>Remarks:</u> Submit electronic request. If you do not have electronic ordering capability, use an ARUP Ai (#32960) with an ARUP client number. For additional technical details, contact ARUP Client Services clinical history. <u>Unacceptable Conditions:</u> Specimens submitted with non-representative tissue type. Specimens that are collection instructions provided in the collection kit. <u>Stability (collection to initiation of testing)</u> : Ambient: 24 hours; Refrigerated: 24 hours; Frozen; Unacce	P supply #40923) available ixatives are required; collection ainer during summer months. natomic Pathology Form at (800) 522-2787. Submit e not collected according to the eptable

Interpretive Data: Refer to report.

Note: Detailed collection instructions are available in the Muscle Biopsy Collection Kit (#40923) or can be requested by contacting ARUP Client Services at (800) 522-2787. Use of a different collection kit could result in sub-optimal biopsy preparation or fixation and delays in diagnosis. All vials must be labeled with the patient's full name and unique identifier. Avoid drying of specimens; make sure the fresh specimens are completely moist with saline gauze and fixed specimen are submerged into the fixative and not caught on vial sides or in the threads of the cap. **Please acquire and use ARUP's Muscle Biopsy Collection Kit as the instructions and contents make the muscle collection procedure safer and easier.**

NOTE: Ancillary testing is ordered at the discretion of the ARUP pathologist, additional CPT codes and charges may apply.

CPT Code(s): 88305, 88313, 88314 x3, 88319 x8

New York DOH Approved.



New Test	<u>2013261</u>	Comprehensive Nerve I	Biopsy Workup		NER REQ
Available Apri	il 18, 2016				
2 	Anatomic Path Recommended	ology Test Request Form (ARUP form #32960)	Ō	Time Sensitive	
Methodology:	Microscopy/Cy	tochemistry			
Performed:	Sun-Sat				
Reported:	1-7 days				
Specimen Requir	red: <u>Collect:</u> Nerve <u>Specimen Prep</u>	biopsy. aration: Obtain Muscle/Nerve Biops	sy collection kit prior to	o collection procedure, (ARUP	supply #40923) available
	instructions are	esupply using ARUP Connect ^{1M} or provided with the kit	contact Client Service	es at $(800)522-2787$. Special ins	catives are required; collection
	Storage/Transp	port Temperature: Room temperature	e. Also acceptable: Ref	frigerated. Ship in cooled conta	iner during summer months.
	Remarks: Subn	nit electronic request. If you do not l	have electronic orderin	ng capability, use an ARUP Ana	atomic Pathology Form
	(#32960) with	an ARUP client number. For addition	onal technical details, c	contact ARUP Client Services a	tt (800) 522-2787. Submit
	Linical history	Conditional Spacimons submitted wi	th non concontative t	issue type. Specimens that are	not collected according to the
	collection instr	uctions provided in the collection ki	iti non-representative t	issue type. Specifiens that are	not conected according to the
	Stability (colled	ction to initiation of testing): Ambie	ent: 24 hours; Refrigera	ated: 24 hours; Frozen; Unacce	otable
Intorprotivo Do	Stability (collected)	ction to initiation of testing): Ambie	ent: 24 hours; Refrigera	ated: 24 hours; Frozen; Unaccep	ptable

Note: Detailed collection instructions are available in the Muscle/Nerve Biopsy Collection Kit (#40923) or can be requested by contacting ARUP Client Services at (800) 522-2787. Use of a different collection kit could result in sub-optimal biopsy preparation or fixation and delays in diagnosis. All vials must be labeled with the patient's full name and unique identifier. Please acquire and use ARUP's Muscle/Nerve Biopsy Collection Kit as the instructions and contents make the muscle/nerve collection procedure safer and easier.

NOTE: Ancillary testing is ordered at the discretion of the ARUP pathologist, additional CPT codes and charges may apply.

CPT Code(s): 88305, 88313, 88348

New York DOH Approved.



New Test Available April	2013257 18, 2016	Consultation, Head and Neck	HN CONSULT
Methodology: Performed:	Microscopy Mon-Fri		

Reported: Varies

Specimen Required: Collect: Tissue.

Specimen Preparation: Formalin fix (10 percent neutral buffered formalin is preferred) and paraffin embed specimen. Protect paraffin block and/or slides from excessive heat. Transport all case material to include stained slides, paraffin blocks and surgical pathology report. Storage/Transport Temperature: Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months. Remarks: Submit electronic request. If you do not have electronic ordering capability, use an ARUP Anatomic Pathology Form (#32960) with an ARUP client number. For additional technical details, contact ARUP Client Services at (800) 522-2787.

Unacceptable Conditions: Specimens submitted with non-representative tissue type. Depleted specimens.

Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable

Interpretive Data: Refer to report.

Note: Appropriate stains and other ancillary testing may be performed and charged separately. Tests requested by the referring physician (eg, immunostains, molecular studies, etc.) may not be performed if they are deemed to be unnecessary by the reviewing ARUP pathologist. For all pathology consultations, ancillary testing is ordered at the discretion of the ARUP pathologist.

CPT Code(s): 88321 or 88323 or 88325, if ancillary testing is performed, additional CPT codes and charges may apply

New York DOH Approved.

HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

New Test	<u>2013262</u>	Consultation, Neuropathology	NP CONSULT				
Available April 18, 2016							
Methodology:	Microscopy						
Performed:	Mon-Fri						
Reported:	Varies						

Specimen Required: Collect: Tissue.

Specimen Preparation: Formalin fix (10 percent neutral buffered formalin is preferred) and paraffin embed specimen. Protect paraffin block and/or slides from excessive heat. Transport all case material to include stained slides, paraffin blocks and surgical pathology report.

Storage/Transport Temperature: Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months. Remarks: Submit electronic request. If you do not have electronic ordering capability, use an ARUP Anatomic Pathology Form (#32960) with an ARUP client number. For additional technical details, contact ARUP Client Services at (800) 522-2787. Unacceptable Conditions: Specimens submitted with non-representative tissue type. Depleted specimens. Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable

Interpretive Data: Refer to report.

Note: Appropriate stains and other ancillary testing may be performed and charged separately. Tests requested by the referring physician (eg immunostains, molecular studies, etc.) may not be performed if they are deemed to be unnecessary by the reviewing ARUP pathologist. For all pathology consultations, ancillary testing is ordered at the discretion of the ARUP pathologist.

CPT Code(s): 88321 or 88323 or 88325, if ancillary testing is performed, additional CPT codes and charges may apply

New York DOH Approved.



New Test	<u>2013263</u>	Consultation, Surgical Pathology	SP CONSULT
Available April	1 18, 2016		
Methodology:	Microscopy		
Performed:	Mon-Fri		
Reported:	Varies		

Specimen Required: Collect: Tissue.

Specimen Preparation: Formalin fix (10 percent neutral buffered formalin is preferred) and paraffin embed specimen. Protect paraffin block and/or slides from excessive heat. Transport all case material to include stained slides, paraffin blocks and surgical pathology report. Storage/Transport Temperature: Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months. Remarks: Submit electronic request. If you do not have electronic ordering capability, use an ARUP Anatomic Pathology Form (#32960) with an ARUP client number. For additional technical details, contact ARUP Client Services at (800) 522-2787.

Unacceptable Conditions: Specimens submitted with non-representative tissue type. Depleted specimens.

Stability (collection to initiation of testing): Ambient: Indefinitely; Refrigerated: Indefinitely; Frozen: Unacceptable

Interpretive Data: Refer to report.

Note: Appropriate stains and other ancillary testing may be performed and charged separately. Tests requested by the referring physician (eg immunostains, molecular studies, etc.) may not be performed if they are deemed to be unnecessary by the reviewing ARUP pathologist. For all pathology consultations, ancillary testing is ordered at the discretion of the ARUP pathologist.

CPT Code(s): 88321 or 88323 or 88325, if ancillary testing is performed, additional CPT codes and charges may apply

New York DOH Approved.

HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

0060360 Corynebacterium diphtheriae Culture

Specimen Required: Collect: Nasopharynx, throat, or wound swab.

Swab nasopharynx and throat at the site of membrane or inflammation to increase recovery. Swab base of cleansed wound, if present. Submit each swab with a separate test order.

0020414 **Creatine Kinase Isoenzymes**

Reference Interval:

Test Number	Components	Reference Interval					
	CK-MM	96-100%					
	CK-MB	0-4%					
	CK-BB	0%					
	CK-Macro Type I	0%					
	CK-Macro Type II	0%					
	Creatine Kinase, Total	Effective May 16, 2016					
		Age: Male: Female:					
		0-30 days	108-564 U/L	108-564 U/L			
		31 days-5 months	72-367 U/L	72-367 U/L			
		6-35 months	50-272 U/L	38-261 U/L			
		3-6 years	56-281 U/L	40-222 U/L			
		7-17 years	60-393 U/L	46-250 U/L			
		18 years and older:	20-200 U/L	20-180 U/L			

MC DIPH

CKISO



2001613 Crohn Disease Prognostic Panel

Interpretive Data: Anti-glycan serologic markers may serve as an aid in the diagnosis of Crohn disease (CD) and an indicator of disease prognosis. Antisaccharomyces cerevaisae antibody (ASCA) has the highest diagnostic value and anti-chitobioside carbohydrate antibody (ACCA) has the highest association with more-aggressive disease. Both ACCA and ASCA are associated equally with the need for surgery. When found in combination, these markers have the best diagnostic potential. A combination of two or more of the four anti-glycan markers have a significant association with complications and the need for surgery than any single marker alone. A negative result does not rule out CD.

See Compliance Statement A: www.aruplab.com/CS

HOT LINE NOTE: There is a component change associated with this test. Add component 2013334, Crohn Disease Prognostic Pan Interp.

New Test	<u>2013294</u>	Dengue Virus (1-4) Subtype by PCR	DENGUEPCR
Available April	18, 2016		

Methodology:	Qualitative Polymerase Chain Reaction
Performed:	Tue, Fri
Reported:	2-5 days

 Specimen Required:
 Collect: Lavender (EDTA), Pink (K2EDTA), or Serum Separator Tube (SST).

 Specimen Preparation:
 Separate serum or plasma from cells. Transfer 1 mL serum or plasma to a sterile container. (Min: 0.5mL)

 Storage/Transport Temperature:
 Frozen.

 Remarks:
 Specimen source required.

 Unacceptable Conditions: Heparinized specimens.
 Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 5 days; Frozen: 2 weeks

Interpretive Data: A negative result does not rule out the presence of PCR inhibitors in the patient specimen or of test-specific nucleic acid in concentrations below the level of detection by this test.

See Compliance Statement B: www.aruplab.com/CS

Note: This assay detects and differentiates Dengue subtypes 1-4.

CPT Code(s): 87798 x4

New York DOH approval pending. Call for status update.

HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

2002247	Disaccharidase, Tissue	DISAC
Performed: Reported:	Mon, Wed, Fri 1-6 days	

Reference Interval: Effective May 16, 2016

Component	Reference Interval
Lactase	Greater than or equal to 10.0 µmol/min/g protein
Maltase	Greater than or equal to 100.0 µmol/min/g protein
Palatinase	Greater than or equal to 9.0 µmol/min/g protein
Sucrase	Greater than or equal to 25.0 µmol/min/g protein

CROHN PAN



2006621 Drug Detection Panel, Umbilical Cord Tissue, Qualitative

TOF SCR CD

Methodology: Qualitative Liquid Chromatography/Tandem Mass Spectrometry/Enzyme-Linked Immunosorbent Assay

Reference Interval: Effective May 16, 2016

Drugs covered and range of cutoff concentrations.

Drugs/Drug Classes	Cutoff Concentrations (ng/g)
Buprenorphine	1
Norbuprenorphine	0.5
Buprenorphine-G	1
Codeine	0.5
Dihydrocodeine	1
Fentanyl	0.5
Hydrocodone	0.5
Norhydrocodone	1
Hydromorphone	0.5
Meperidine	2
Methadone	2
Methadone metabolite	1
6-Acetylmorphine	1
Morphine	0.5
Naloxone	1
Oxycodone	0.5
Noroxycodone	1
Oxymorphone	0.5
Noroxymorphone	0.5
Propoxyphene	1
Tapentadol	2
Tramadol	2
N-desmethyltramadol	2
O-desmethyltramadol	2
Amphetamine	5
Benzoylecgonine	0.5
m-OH-Benzoylecgonine	1
Cocaethylene	1
Cocaine	0.5
MDMA (Ecstasy)	5
Methamphetamine	5
Phentermine	8
Alprazolam	0.5
Alpha-OH-Alprazolam	0.5
Butalbital	25
Clonazepam	1
7-Aminoclonazepam	
Diazepam	
Lorazepam	5
Midazolam	
Alpha-OH-Midazolam	2
Nordiazepam	
Uxazepam	
Phenobarbital	15
Temazepam	
	0.5
Phencyclidine (PCP)	1
Marijuana Metabolite	1

Interpretive Data:

Methodology: Qualitative Liquid Chromatography/Tandem Mass Spectrometry/Enzyme-Linked Immunosorbent Assay

Detection of drugs in umbilical cord tissue is intended to reflect maternal drug use during pregnancy. The pattern and frequency of drug(s) used by the mother cannot be determined by this test. A negative result does not exclude the possibility that a mother used drugs during pregnancy. Detection of drugs in umbilical cord tissue depends on extent of maternal drug use, as well as drug stability, unique characteristics of drug deposition in umbilical cord tissue, and the performance of the analytical method. Drugs administered during labor and delivery may be detected. Detection of drugs in umbilical cord tissue does not insinuate impairment and may not affect outcomes for the infant. Interpretive questions should be directed to the laboratory. Glucuronide metabolites are indicated as -G.

For medical purposes only; not valid for forensic use unless testing was performed within Chain of Custody process. See Compliance Statement B: www.aruplab.com/CS



HOT LINE NOTE: There is a component change, unit of measure change and clinically significant charting name change associated with this test. Remove component 2006623, OPIOIDS, UMBILICAL CORD Remove component 2006624, STIMULANTS, UMBILICAL CORD Remove component 2006625, SEDATIVES-HYPNOTICS, UMBILICAL CORD Remove component 2006684, PHENCYCLIDINE, UMBILICAL CORD Remove component 2008359, OTHER, IMMUNOASSAY, UMBILICAL CORD Change the charting name of component 2006622 from Drug Detection Pan, TOF, Umbilical Cord to Drug Detection Panel, Umbilical Cord Change the charting name of component 2006626 from Buprenorphine (cutoff 2 ng/g) to Buprenorphine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2013103 from Norbuprenorphine (cutoff 8 ng/g) to Norbuprenorphine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006627 from Buprenorphine-G (cutoff 8 ng/g) to Buprenorphine-G, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006628 from Codeine (cutoff 6 ng/g) to Codeine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006629 from Dihydrocodeine (cutoff 4 ng/g) to Dihydrocodeine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006630 from Fentanyl (cutoff 1 ng/g) to Fentanyl, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006631 from Hydrocodone (cutoff 6 ng/g) to Hydrocodone, Cord, Qual and add unit of measure ng/g Change the charting name of component 2013104 from Norhydrocodone (cutoff 6 ng/g) to Norhydrocodone, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006632 from Hydromorphone (cutoff 4 ng/g) to Hydromorphone, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006633 from Meperidine (cutoff 2 ng/g) to Meperidine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006634 from Methadone (cutoff 10 ng/g) to Methadone, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006635 from EDDP (cutoff 10 ng/g) to Methadone Metabolite, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006636 from 6-Acetylmorphine (cutoff 4 ng/g) to 6-Acetylmorphine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006637 from Morphine (cutoff 4 ng/g) to Morphine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006638 from Naloxone (cutoff 8 ng/g) to Naloxone, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006640 from Oxycodone (cutoff 4 ng/g) to Oxycodone, Cord, Qual and add unit of measure ng/g Change the charting name of component 2013105 from Noroxycodone (cutoff 4 ng/g) to Noroxycodone, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006641 from Oxymorphone (cutoff 4 ng/g) to Oxymorphone, Cord, Qual and add unit of measure ng/g Change the charting name of component 2013106 from Noroxymorphone (cutoff 4 ng/g) to Noroxymorphone, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006642 from Proposyphene (cutoff 10 ng/g) to Proposyphene, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006644 from Tapentadol (cutoff 2 ng/g) to Tapentadol, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006645 from Tramadol (cutoff 2 ng/g) to Tramadol, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006646 from N-desmethyltramadol (cutoff 2 ng/g) to N-desmethyltramadol, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006647 from O-desmethyltramadol (cutoff 2 ng/g) to O-desmethyltramadol, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006648 from Amphetamine (cutoff 8 ng/g) to Amphetamine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006649 from Benzoylecgonine (cutoff 8 ng/g) to Benzoylecgonine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006650 from m-OH-Benzoylecgonine (cutoff 8 ng/g) to m-OH-Benzoylecgonine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006652 from Cocaethylene (cutoff 8 ng/g) to Cocaethylene, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006653 from Cocaine (cutoff 8 ng/g) to Cocaine, Cord, Qual and add unit of measure ng/g

Change the charting name of component 2006654 from MDMA- Ecstasy (cutoff 8 ng/g) to MDMA - Ecstasy, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006657 from Methamphetamine (cutoff 8 ng/g) to Methamphetamine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006658 from Phentermine (cutoff 8 ng/g) to Phentermine, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006659 from Alprazolam (cutoff 5 ng/g) to Alprazolam, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006660 from Alpha-OH-Alprazolam (cutoff 5 ng/g) to Alpha-OH-Alprazolam, Cord, Qual and add unit of measure ng/g

Change the charting name of component 2006662 from Butalbital (cutoff 75 ng/g) to Butalbital, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006663 from Clonazepam (cutoff 5 ng/g) to Clonazepam, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006664 from 7-Aminoclonazepam (cutoff 5 ng/g) to 7-Aminoclonazepam, Cord, Qual and add unit of measure ng/g

Change the charting name of component 2006665 from Diazepam (cutoff 5 ng/g) to Diazepam, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006671 from Lorazepam (cutoff 5 ng/g) to Lorazepam, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006672 from Midazolam (cutoff 5 ng/g) to Midazolam, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006673 from Alpha-OH-Midazolam (cutoff 5 ng/g) to Alpha-OH-Midazolam, Cord, Qual and add unit of measure ng/g

Change the charting name of component 2006675 from Nordiazepam (cutoff 5 ng/g) to Nordiazepam, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006676 from Oxazepam (cutoff 5 ng/g) to Oxazepam, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006677 from Phenobarbital (cutoff 75 ng/g) to Phenobarbital, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006679 from Temazepam (cutoff 5 ng/g) to Temazepam, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006682 from Zolpidem (cutoff 10 ng/g) to Zolpidem, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006682 from Zolpidem (cutoff 10 ng/g) to Zolpidem, Cord, Qual and add unit of measure ng/g Change the charting name of component 2006683 from Phencyclidine- PCP (cutoff 4 ng/g) to Phencyclidine - PCP, Cord, Qual and add unit of measure ng/g Change the charting name of component 2008360 from Marijuana Metabolite (cutoff 1 ng/g) to Marijuana Metabolite, Cord, Qual and add unit of measure ng/g



2008603 ERBB2 (HER2/neu) Gene Amplification by FISH, Tissue

ERBB2 FISH

Specimen Required: Specimen Preparation: Formalin fix (10 percent neutral buffered formalin) and paraffin-embed tissue. Fixative duration: 6-72 hours. Protect paraffin block from excessive heat. Transport block or 5 unstained (4-micron thick sections) positively charged slides in a tissue transport kit (ARUP supply #47808) available online through eSupply using ARUP Connect™ or contact ARUP Client Services at (800) 522-2787 (kit recommended but not necessary). (Min 2 slides) Unacceptable Conditions: Specimens fixed or processed in alternative fixatives (alcohol, Prefer) or heavy metal fixatives (B-4 or B-5). Tissue fixed for less than 6 hours or greater than 72 hours. No tumor in tissue. Decalcified specimens.

New Test2013277Esterase, Non-Specific Cytochemical Stain OnlyAvailable May 16, 2016

NSE SO

ETG SCR UR

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Time Sensitive

Methodology:	Cytochemical Stain
Performed:	Mon-Fri
Reported:	2-5 days

Specimen Required: Collect: Lavender (EDTA), Green (Lithium Heparin), or Green (Sodium Heparin). Also acceptable: Heparinized Bone Marrow Aspirate. Specimen Preparation: Blood: Protect from light. Transport 5 mL whole blood AND 6 unfixed, air-dried, and unstained push

smears made from the blood submitted. (Min: 1 mL AND 6 unfixed smears). OR Bone Marrow: Protect from light. Transport 1 mL heparinized aspirate AND 6 unfixed, air-dried, and unstained bone marrow aspirate smears. (Min: 0.5 mL AND 6 unfixed smears).

Storage/Transport Temperature: Room temperature. Specimens should be **received** within 24 hours of collection; testing must be **performed** within 48 hours of collection.

<u>Unacceptable Conditions:</u> Peripheral blood smears older than one year. Anticoagulated blood or bone marrow sent without wellprepared unfixed smears older than 48 hours.

Stability (collection to initiation of testing): Ambient: 48 hours; Refrigerated: Unacceptable; Frozen: Unacceptable

Note: Further information on how to make an adequate slide, in the form of an instructional video, can be found at: https://www.youtube.com/watch?v=ca3NwrlpS40&feature=youtube

CPT Code(s): 88319

HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

<u>2012695</u> Ethyl Glucuronide Screen Only, Urine

Reference Interval: Screen cutoff concentration: 500 ng/mL



0040080

Hematocrit

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НСТ

Time Sensitive

Specimen Required: Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 48 hours; Frozen: Unacceptable

Reference Interval: Effective May 16, 2016

Age	0 days	1-6 days	7-13 days	14-29 days	30-60 days	61-90 days	91-180 days	6-23 months	2-5 years	6-11 years	12-17 years	18 years and older
Male %	42-60%	45-64.9%	42-64.9%	39-63%	31-55%	28-42%	29-41%	33-39%	34-40%	35-45%	37-49%	44.2-53%
Female %	42-60%	45-64.9%	42-64.9%	39-63%	31-55%	28-42%	29-41%	33-39%	34-40%	35-45%	36-46%	36-49%

<u>0040085</u>

Hemoglobin



Time Sensitive

Specimen Required:

Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 48 hours; Frozen: Unacceptable

Reference Interval: Effective May 16, 2016

Age	0 days	1-6 days	7-13 days	14-29 days	30-60 days	61-90 days	91-180 days	6-23 months	2-5 years	6-11 years	12-17 years	18 years and older
Male (g/dL)	13.5-19.5	14.5-22.5	13.5-21.5	12.5-20.5	10.0-18.0	9.0-14.0	9.5-13.5	10.5-13.5	11.5-13.5	11.5-15.5	13.0-16.0	14.8-17.8
Female (g/dL)	13.5-19.5	14.5-22.5	13.5-21.5	12.5-20.5	10.0-18.0	9.0-14.0	9.5-13.5	10.5-13.5	11.5-13.5	11.5-15.5	12.0-16.0	12.6-15.9

0020416 Hepatic Function Panel

HOT LINE NOTE: There is a unit of measure change associated with this test. Change unit of measure for component 0020007, Aspartate Aminotransferase from IU/L to U/L

Change unit of measure for component 0020008, Alanine Aminotransferase from IU/L to U/L

0092522 Histoplasma Antigen by EIA, Serum

Specimen Required: Stability (collection to initiation of testing): Ambient: Unacceptable; Refrigerated: 1 week; Frozen: 1 month (avoid repeated freeze/thaw cycles)

HISTOAG S

HEPATIC

HGB



New Test2013333Human Immunodeficiency Virus (HIV) Combo
Antigen/Antibody (HIV-1/O/2) by ELISA, with Reflex to HIV-
1/HIV-2 Antibody Differentiation, Supplemental

HIVAGABGE

Available April 18, 2016

Methodology:	Qualitative Enzyme-Linked Immunosorbent Assay/Qualitative Immunoassay
Performed:	Mon, Wed-Sat
Reported:	1-3 days

 Specimen Required: Collect: Serum Separator Tube (SST). Also acceptable: Green (Sodium or Lithium Heparin), Lavender (EDTA), Pink (K2EDTA), Red (Clot Activator), or Plasma Separator Tube (PST).

 Specimen Preparation: Separate from cells ASAP or within 2 hours of collection. Transfer 1 mL serum into an ARUP Standard Transport Tube. (Min: 0.5 mL) Remove particulate material.

 Storage/Transport Temperature: Refrigerated.

 Unacceptable Conditions: Specimens containing particulate material. Severely hemolyzed or heat-inactivated specimens.

 Stability (collection to initiation of testing): After separation from cells: Ambient: Unacceptable; Refrigerated: 1 week; Frozen: Indefinitely (avoid repeated freeze/thaw cycles)

Reference Interval:

Test Number	Components	Reference Interval		
	HIV 1,2 Combo	Negative		
	Antigen/Antibody			
2013107	Human Immunodeficiency Virus Types 1 and 2 (HIV-1/2) Antibody Differentiation, Supplemental			
		Available Separately	Components	Reference Interval
		No	HIV-1 Antibody	Negative
		No	HIV-2 Antibody	Negative

Interpretive Data: This test should not be used for blood donor screening, associated re-entry protocols, or for screening Human Cell, Tissues and Cellular and Tissue-Based Products (HCT/P).

Note: The fourth-generation ELISA screen test is for the simultaneous qualitative detection of Human Immunodeficiency Virus Type 1 (HIV-1) p24 antigen and antibodies to HIV Type 1 (HIV-1 groups M and O) and HIV Type 2 (HIV-2). Results of the screen cannot be used to distinguish between the presence of HIV-1 p24 antigen, HIV-1 antibody, or HIV-2 antibody.

The reflexed HIV-1/HIV-2 Antibody Differentiation test discriminates between HIV-1 and HIV-2 antibodies. Results for each type are reported.

If the HIV-1,2 Combo Antigen/Antibody screen is repeatedly reactive, then the HIV-1/ HIV-2 Antibody Differentiation test will be performed. Additional charges apply. A recommendation to order further testing on a separate specimen for HIV-1 Nucleic Acid will be made for certain results. This multi-test algorithm was proposed by the Centers for Disease Control and Prevention (CDC) and adopted by the Clinical Laboratory Standards Institute (CLSI) for the diagnosis of HIV.

CPT Code(s): 87389; if reflexed, add 86701 and 86702

New York DOH approved.

HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

2002899 Human Papillomavirus (HPV), High Risk by in situ Hybridization, Paraffin

HPVHI ISH

HOT LINE NOTE: There is a component change associated with this test. Change component 2003001 H and E Slide Description from a Prompt test to a Resultable test.



0050980 Humoral Immunity Panel I

HOT LINE NOTE: There is a clinically significant charting name change associated with this test. Change the charting name of component 0050706 from Pneumococcal Serotype 1 IgG to Pneumo serotype 1 IgG (P13,PNX) Change the charting name of component 0050709 from Pneumococcal Serotype 4* IgG to Pneumo serotype 4 IgG (P7,P13,PNX) Change the charting name of component 0050709 from Pneumococcal Serotype 5 IgG to Pneumo serotype 5 IgG (P13,PNX) Change the charting name of component 0050713 from Pneumococcal Serotype 6B* IgG to Pneumo serotype 6B IgG (P7,P13,PNX) Change the charting name of component 0050715 from Pneumococcal Serotype 3 IgG to Pneumo serotype 3 IgG (P13,PNX) Change the charting name of component 0050716 from Pneumococcal Serotype 7F IgG to Pneumo serotype 7F IgG (P13,PNX) Change the charting name of component 0050717 from Pneumococcal Serotype 9N IgG to Pneumo serotype 9N IgG (PX) Change the charting name of component 0050717 from Pneumococcal Serotype 9N IgG to Pneumo serotype 9N IgG (P7,P13,PNX) Change the charting name of component 0050712 from Pneumococcal Serotype 14* IgG to Pneumo serotype 14 IgG (P7,P13,PNX) Change the charting name of component 0050721 from Pneumococcal Serotype 8 IgG to Pneumo serotype 8 IgG (PX) Change the charting name of component 0050722 from Pneumococcal Serotype 9V*, IgG to Pneumo serotype 9V IgG (P7,P13,PNX) Change the charting name of component 0050723 from Pneumococcal Serotype 12F IgG to Pneumo serotype 12F IgG (PX) Change the charting name of component 0050724 from Pneumococcal Serotype 12F IgG to Pneumo serotype 18C IgG (P7,P13,PNX) Change the charting name of component 0050724 from Pneumococcal Serotype 18C* IgG to Pneumo serotype 18C IgG (P7,P13,PNX) Change the charting name of component 0050726 from Pneumococcal Serotype 19F* IgG to Pneumo serotype 19F IgG (P7,P13,PNX) Change the charting name of component 0050726 from Pneumococcal Serotype 19F* IgG to Pneumo serotype 18C IgG (P7,P13,PNX) Change the charting name of component 0050726 from Pneumococcal Serotyp

0050981 Humoral Immunity Panel II

HUMPAN II

HOT LINE NOTE: There is a clinically significant charting name change associated with this test. Change the charting name of component 0050706 from Pneumococcal Serotype 1 IgG to Pneumo serotype 1 IgG (P13,PNX) Change the charting name of component 0050709 from Pneumococcal Serotype 4* IgG to Pneumo serotype 4 IgG (P7,P13,PNX) Change the charting name of component 0050709 from Pneumococcal Serotype 5 IgG to Pneumo serotype 5 IgG (P13,PNX) Change the charting name of component 0050713 from Pneumococcal Serotype 6B* IgG to Pneumo serotype 6B IgG (P7,P13,PNX) Change the charting name of component 0050715 from Pneumococcal Serotype 3 IgG to Pneumo serotype 3 IgG (P13,PNX) Change the charting name of component 0050716 from Pneumococcal Serotype 7F IgG to Pneumo serotype 7F IgG (P13,PNX) Change the charting name of component 0050717 from Pneumococcal Serotype 7N IgG to Pneumo serotype 9N IgG (PNX) Change the charting name of component 0050718 from Pneumococcal Serotype 14* IgG to Pneumo serotype 14 IgG (P7,P13,PNX) Change the charting name of component 0050712 from Pneumococcal Serotype 14* IgG to Pneumo serotype 8 IgG (PX) Change the charting name of component 0050721 from Pneumococcal Serotype 8V; IgG to Pneumo serotype 8 IgG (PX) Change the charting name of component 0050722 from Pneumococcal Serotype 9V*, IgG to Pneumo serotype 8 IgG (PX) Change the charting name of component 0050724 from Pneumococcal Serotype 12F IgG to Pneumo serotype 12F IgG (PX) Change the charting name of component 0050724 from Pneumococcal Serotype 18K² IgG to Pneumo serotype 18C IgG (P7,P13,PNX) Change the charting name of component 0050724 from Pneumococcal Serotype 18K² IgG to Pneumo serotype 19F IgG (P7,P13,PNX) Change the charting name of component 0050724 from Pneumococcal Serotype 18K² IgG to Pneumo serotype 19F IgG (P7,P13,PNX) Change the charting name of component 0050724 from Pneumococcal Serotype 19F* IgG to Pneumo serotype 19F IgG (P7,P13,PNX) Change the charting name of component 0050726 from Pneumococcal Serotype 19F*



New Test	<u>2013270</u>	Inflammatory Bowel Disease Differentiation Panel	IBD PAN		
Available July	5, 2016				
Methodology:	Semi-Quantitat	ive Enzyme-Linked Immunosorbent Assay/Semi-Quantitative Indirect Fluorescent Antibo	ody		
Performed:	Refer to individ	Refer to individual components			
Reported:	1-4 days				
Specimen Require	ed: Collect: Serum	Separator Tube (SST).			
	Specimen Prepa	aration: Separate from cells ASAP or within 2 hours of collection. Transfer 1.5 mL serum	to an ARUP Standard		
	Transport Tube	. (Min: 0.6 mL)			
	Storage/Transp	ort Temperature: Refrigerated.			
	Unacceptable C	conditions: Contaminated, heat-inactivated, hemolyzed, or severely lipemic specimens.			

<u>Stability (collection to initiation of testing)</u>: After separation from cells: Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year (avoid repeated freeze/thaw cycles)

Reference Interval:

Test Number	Components	Reference Interval
	Saccharomyces cerevisiae	20.0 Units or less: Negative
	Antibody, IgG	20.1-24.9 Units: Equivocal 25.0 Units or greater: Positive
	Saccharomyces cerevisiae	20.0 Units or less: Negative
	Antibody, IgA	20.1-24.9 Units: Equivocal
		25.0 Units or greater: Positive
0050811	Anti-Neutrophil	Less than 1:20: Not significant
	Cytoplasmic Antibody,	
	IgG	

Interpretive Data: Refer to report.

Note: This test may be a useful tool for distinguishing ulcerative colitis (UC) from Crohn disease (CD) in patients with suspected inflammatory bowel disease. If the ANCA screen detects antibodies at a 1:20 dilution or greater, then a titer to end point will be added. Additional charges apply. ANCA IFA is simultaneously tested on ethanol- and formalin-fixed slides to allow differentiation of C- and P-ANCA patterns.

CPT Code(s): 86671 x2; 86255; if reflexed, add 86256

New York DOH Approved.

HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

<u>0049110</u>	Iron Stain	IRON STAIN
Specimen Requi	red: <u>Collect:</u> Lavender (EDTA) bone marrow aspirate. <u>Specimen Preparation:</u> Transfer 4 unfixed, air-dried, unstained, non-anticoagulated bone marrow touch preps to a metal free container. (Min: 1 mL) <u>Unacceptable Conditions:</u> Peripheral blood. Fixed smears.	w aspirate, EDTA smears, or core
0080200	Lecithin-Sphingomyelin Ratio	L-S
Specimen Requi	red: <u>Unacceptable Conditions</u> : Specimens grossly contaminated with blood, mucus, meconium, or u interference.	rine, are unacceptable due to

FIBRO V

2005661 Liver Fibrosis, Chronic Viral Hepatitis (Echosens FibroMeter)

HOT LINE NOTE: There is a unit of measure change associated with this test.

Change unit of measure for component 2010928, Aspartate Aminotransferase, FibroMeter from IU/L to U/L Change unit of measure for component 2010929, Alanine Aminotransferase, FibroMeter from IU/L to U/L



0040005 Manual Differential

DIFF

Reference Interval: Effective May 16, 2016

Test Number	Components	Reference Interval					
	PMNs	12 yrs & over	12 yrs & over 44 – 76%				
	Bands	0-5%					
	Lymphocytes	12 yrs & over	15 – 43%				
	Monocytes	12 yrs & over	2 – 8%				
	Eosinophil %	0-6%					
	Basophil Percentage	12 yrs & over	12 yrs & over 0 – 2%				
	Absolute Neutrophil Count	Age	0-11 months	1-5 years	6-13 years	14-17 years	18 years and older
		K/µL	1.5-10.0	1.5-8.5	1.5-8.0	1.8-8.0	2.0-7.4
	Morphology	Normocytic/N	ormochromic				
	Platelet (estimate)	Adequate					



Reference Interval: Not Detected

Interpretive Data: The meningitis/encephalitis panel is NOT a replacement for CSF bacterial and/or fungal culture and Cryptococcal antigen testing for at-risk patients. Non-K1 *E. coli* serotypes and non-encapsulated strains of *Neisseria meningitidis* are NOT detected. The panel does NOT differentiate active from latent herpes virus infections. Test results should be interpreted in the context of host factors and other laboratory information.

Note: This panel includes *Cryptococcus neoformans/gattii*, Cytomegalovirus (CMV), Enterovirus, *Escherichia coli* K1, *Haemophilus influenzae*, Herpes simplex virus 1(HSV-1), Herpes simplex virus 2(HSV-2), Human herpesvirus 6 (HHV-6), Human parechovirus, *Listeria monocytogenes*, *Neisseria meningitidis*, *Streptococcus agalactiae*, *Streptococcus pneumoniae*, Varicella-zoster virus (VZV).

CPT Code(s): 87496; 87498; 87529 x2; 87532; 87653; 87798 x8

New York DOH Approved.

HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

MS SEQ-1	
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Specimen Required: <u>Patient Prep</u>: This test requires a nuchal translucency (NT) measurement that has been performed by a certified ultrasonographer. The ultrasonographer MUST be certified to perform NT measurements by one of the following agencies: Fetal Medicine Foundation (FMF) or Nuchal Translucency Quality Review (NTQR). To avoid possible test delays for an ultrasonographer that is new to our database, please contact the genetic counselor at 800-242-2787 extension 2141 prior to sending specimen.

If an NT is unobtainable, order Maternal Serum Screening, Integrated (ARUP test codes 0081062 and 0081064), which can be interpreted without an NT value.

Specimen must be drawn between 11 weeks, 0 days and 13 weeks, 6 days gestation (Crown-Rump length (CRL) must be 4.4-8.5 cm).

Specimen Required: <u>Patient Prep</u>: This test requires a nuchal translucency (NT) measurement that has been performed by a certified ultrasonographer. The ultrasonographer MUST be certified to perform NT measurements by one of the following agencies: Fetal Medicine Foundation (FMF) or Nuchal Translucency Quality Review (NTQR). To avoid possible test delays for an ultrasonographer who is new to our database, please contact the genetic counselor at 800-242-2787 extension 2141 prior to sending specimen.

If an NT is unobtainable, order Maternal Serum Screening, Integrated (ARUP test codes 0081062 and 0081064), which can be interpreted without an NT value.

Specimen must be drawn in the first trimester between 11 weeks, 0 days and 13 weeks, 6 days. (Crown-Rump length (CRL) must be between 4.4-8.5 cm). Patient History information is required

0081062 Maternal Serum Screening, Integrated, Specimen #1

Maternal Screening, Sequential, Specimen #1

Specimen Required: <u>Patient Prep:</u> The nuchal translucency (NT) measurement is preferred; however, the Integrated Maternal Screen can be interpreted with or without a NT measurement. If performed, the NT measurement must be obtained between 10 weeks, 3 days and 13 weeks, 6 days gestation (Crown-Rump length (CRL) must be 3.9-8.5 cm). The NT measurement must also be performed by an ultrasonographer that is certified by one of the following agencies: Fetal Medicine Foundation (FMF) or Nuchal Translucency Quality Review (NTQR). To avoid possible test delays for an ultrasonographer that is new to our database, please contact the genetic counselor at (800) 242-2787 extension 2141 prior to sending specimen.

Serum-only specimens may be drawn between 10 weeks, 0 days and 13 weeks, 6 days gestation. CRL must be 3.4-8.5 cm. The specimen collection and ultrasound date may be different.

New Test	<u>2013305</u>	Meningitis/Encephalitis Panel by PCR	MEFAP
Available April 1	8, 2016		

Methodology:	Qualitative Polymerase Chain Reaction
Performed:	Sun-Sat
Reported:	Within 24 hours

Specimen Required: Collect: CSF.

0081293

Specimen Preparation: Transfer 0.5 mL CSF to a sterile ARUP Standard Transport Tube (ARUP supply #43115) available online through eSupply using ARUP Connect or contact ARUP Client Services at (800) 522-2787. (Min: 0.25 mL) Do not centrifuge. Storage/Transport Temperature: Refrigerated. Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 1 week; Frozen: Unacceptable

MS FT

MS INT-1



2011521 Meprobamate, Serum or Plasma, Quantitative

MEPRO SP

Reference Interval: Effective May 16, 2016

Dose-Related Range	5.0-20.0 μg/mL
Toxic	Greater than 40.0 µg/mL

HOT LINE NOTE: There is a numeric map change associated with this test.

Change the numeric map for component 2011456, Meprobamate, Serum or Plasma from XXXX to XXXX.X

New Test	<u>2013273</u>	Myeloperoxidase, Cytochemical Stain Only	MPO SO
Available May	16, 2016		

Methodology:	Cytochemical Stain
Performed:	Mon-Fri
Reported:	2-5 days

Specimen Required: <u>Collect:</u> Lavender (EDTA), Green (Lithium Heparin), or Green (Sodium Heparin). Also acceptable: Heparinized Bone Marrow Aspirate.

Specimen Preparation: Transport 6 unfixed, air-dried, and unstained push smears **AND** 5 mL whole blood (smears should be made from the blood submitted). **OR** 6 unfixed, air-dried, and unstained bone marrow aspirate smears. <u>Storage/Transport Temperature</u>: Room temperature. Blood specimens must be **received** within 24 hours of collection; testing must be **performed** within 48 hours of collection. <u>Unacceptable Conditions</u>: Peripheral blood smears older than one week. Anticoagulated blood or bone marrow sent without wellprepared unfixed smears if greater than 48 hours old. <u>Stability (collection to initiation of testing)</u>: Ambient: 24 hours; Refrigerated: Unacceptable; Frozen: Unacceptable

Note: Further information on how to make an adequate slide, in the form of an instructional video, can be found at: https://www.youtube.com/watch?v=ca3NwrlpS40&feature=youtube

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CPT Code(s): 88319
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HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

<u>2008767</u>	Opioid Receptor, mu OPRM1 Genotype, 1 Variant	OPRM1

Methodology:	Polymerase Chain Reaction/Fluorescence Monitoring
Performed:	Mon, Thu
Reported:	5-10 days

 Specimen Required: Collect: Lavender (EDTA), Pink (K2EDTA), or Yellow (ACD Solution A or B).

 Unacceptable Conditions: Plasma or serum. Heparinized specimens.

 Stability (collection to initiation of testing): Ambient: 72 hours; Refrigerated: 2 weeks; Frozen: 1 month

Interpretive Data:

Background Information for Opioid Receptor, Mu OPRM1 Genotype, 1 Variant:

Characteristics: The mu opioid receptor is involved in mediating the clinical response to opioids (agonists and antagonists). *OPRM1* c.118A>G has been associated with lower sensitivity to opioid receptor agonists prescribed for pain control (eg., morphine) and higher sensitivity to opioid receptor antagonists used in the treatment of alcohol and drug dependency (eg., naltrexone). Risk of side effects to opioids is also associated with this genetic variant. **Inheritance:** Autosomal co-dominant.

Cause: SNP rs1799971; OPRM1 c.118A>G (p.Asn40Asp); also known as G allele alters response to opioids.

G allele frequency: African Americans 4 percent, Caucasians 14 percent, Hispanics 24 percent.

Clinical Sensitivity: Drug dependent.

Methodology: Polymerase Chain Reaction (PCR) and Fluorescence Monitoring

Analytical Sensitivity and Specificity: Greater than 99 percent.

Limitations: Only the targeted *OPRM1* mutation, c.118A>G, will be detected. Diagnostic errors can occur due to rare sequence variations. Risk of therapeutic failure or adverse reactions with opioids may be affected by genetic and non-genetic factors that are not detected by this test. This result does not replace the need for therapeutic or clinical monitoring.

See Compliance Statement C: www.aruplab.com/CS



2007479	Pain Management Drug Panel by High-Resolution Time-of-Flight Mass	PAIN HYB U
	Spectrometry and Enzyme Immunoassay, Urine	
HOT LINE NO	TE: There is a component change associated with this test.	
Remove compone	nt 2007667, Ritalinic Acid (cutoff 400 ng/mL) 013281, Methylphenidate (cutoff 100 ng/mL)	
Add component 2	015201, Methylphendale (euton 100 hg/m2)	
<u>2009288</u>	Pain Management Drug Screen with Interpretation by High-Resolution Time- of-Flight Mass Spectrometry and Enzyme Immunoassay, Urine	PAIN HYB 2
HOT LINE NO	TE: There is a component change associated with this test.	
Remove compone	nt 2007667, Ritalinic Acid (cutoff 400 ng/mL)	
Add component 2	013281, Methylphenidate (cutoff 100 ng/mL)	
2012603	PAX8-PPARG Translocations Detection by PCR	PAX8-PPARG
Performed:	RNA isolation: Sun-Sat	
	Assay: Mon, Thu	
Reported:	10-12 days	
0091260	Phenol Exposure Quantitative, Urine	PHENOL U
Performed:	Varies	
Reported:	4-11 days	
<u>2010481</u>	Phenytoin, Free	DIL FR
Reference Inter	rval: Effective May 16, 2016	

Therapeutic:	1.0-2.5 μg/mL
Toxic:	Greater than 2.5µg/mL

0090141 Phenytoin, Free and Total

Reference Interval: Effective May 16, 2016

Available Separately	Components	Therapeutic Range
No	Phenytoin - Total	Therapeutic: 10.0-20.0
		Toxic: > 30.0
No	Phenytoin - Free Level	Therapeutic: 1.0-2.5
		Toxic: > 2.5
No	Phenytoin - Percent Free	8.0-14.0%

0040235

Reference Interval: Effective May 16, 2016

Platelets

Age Range			Sex	Normal Low	Normal High	Units	
0	Days	150	Years	Male/Female	159	439	k/uL

2003040 PM/Scl-100 Antibody, IgG by Immunoblot with Reflex to ANA IFA

HOT LINE NOTE: There is a price change associated with this test. Please contact ARUP Client Services at (800) 522-2787 for additional information.

2002871 *PML-RARA* Translocation, t(15;17) by RT-PCR, Quantitative

Interpretive Data: Refer to report.

See Compliance Statement B: www.aruplab.com/CS

PLT

FDIL

PM/SCL

PML QNT



0095044 **Prenatal Reflexive Panel** PRENATAL A

Automated Cell Count/Differential/Semi-Quantitative Charcoal Agglutination/Qualitative Chemiluminescent Immunoassay/Semi-Quantitative Chemiluminescent Immunoassay/Hemagglutination/Solid Phase Methodology:

2008509 Progesterone Quantitative by HPLC-MS/MS, Serum or Plasma

PGSN

Reference Interval: Effective May 16, 2016

Age	Males
Less than 1 year	Not Established
1-16 years	Less than or equal to 0.15 ng/mL
17 years and older	Less than or equal to 0.11 ng/mL

Age	Females
Less than 1 year	Not Established
1-10 years	Less than or equal to 0.26 ng/mL
11 years	Less than or equal to 2.55 ng/mL
12 years	Less than or equal to 8.56 ng/mL
13 years	Less than or equal to 6.93 ng/mL
14 years	Less than or equal to 12.04 ng/mL
15 years	Less than or equal to 10.76 ng/mL
16 years	Less than or equal to 12.94 ng/mL
17 years and older	Based on Cycle Days
1-6 days	Less than or equal to 0.17 ng/mL
7-12 days	Less than or equal to 1.35 ng/mL
13-15 days	Less than or equal to 15.63 ng/mL
16-28 days	Less than or equal to 25.55 ng/mL
Post-Menopausal	Less than or equal to 0.10 ng/mL
Pregnancy, First Trimester	6.25 – 45.46 ng/mL
Pregnancy, Second Trimester	15.40 – 52.10 ng/mL
Pregnancy, Third Trimester	24.99 – 99.92 ng/mL



New Test	<u>2013352</u>	Pyridoxine-Dependent Epilepsy Panel, Serum or Plasma	P-DEP SP
Available May	16, 2016		
	Patient History	For Biochemical Genetics	
Methodology:	Quantitative L	iquid Chromatography-Tandem Mass Spectrometry	
Performed:	Fri		
Reported:	4-12 days		
Specimen Require	ed: Patient Prep: A	Adults: Fasting specimen preferred.	
	Infants and Ch	ildren: Draw specimen prior to feeding or 2-3 hours after a meal preferred	
	Collect: Green	(Sodium or Lithium Heparin), Lavender (EDTA), Plain Red, or Serum Separator Tube (SST	Г).
	Specimen Prep	paration: Separate from cells ASAP or within 2 hours of collection. Transfer 0.5 mL serum o	r plasma to an ARUP
	Standard Trans	sport Tube and freeze immediately. (Min: 0.2 mL)	
	Storage/Transp	port Temperature: CRITICAL FROZEN. Separate specimens must be submitted when multip	ble tests are ordered.
	Remarks: Clin	ical information is needed for appropriate interpretation. Submit age, gender, diet (eg., TPN	therapy), drug therapy, and
	family history	on a Biochemical Genetics Patient History Form available at www.aruplab.com/patienthisto	ry or by contacting ARUP
	Unaccontable	S at (800) 522-2787.	
	Stability (colle	<u>Conditions</u> : Samples exposed to more than two freeze/fnaw cycles.	ted: 24 hours: Frozen at
	20°C · 1 week	Frozen at -70° C · 1 year	icu. 24 nours, 110201 at -
	20 C. 1 WEEK,	11020h ut 70 C. 1 you	

Reference Interval:

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Age	Pipecolic Acid	Total AASA-P6C		
0-12 months	Less than or equal to 5.2 umol/L	Less than or equal to 1.6 umol/L		
Greater than 1 year	Less than or equal to 6.3 umol/L	Less than or equal to 3.1 umol/L		

Interpretive Data: Refer to report

See Compliance Statement B: www.aruplab.com/CS

CPT Code(s): 82542

New York DOH approval pending. Call for status update.



New Test	<u>2013355</u>	Pyridoxine-Dependent Epilepsy Panel, Urine	P-DEP U
Available May	7 16, 2016		
	Patient History	For Biochemical Genetics	
Methodology: Performed: Reported:	Quantitative L Fri 4-12 days	iquid Chromatography-Tandem Mass Spectrometry	
Specimen Requir	red: <u>Collect:</u> Rande <u>Specimen Prep</u> <u>Storage/Trans</u> 24 hours from <u>Remarks:</u> Clin (e.g. TPN thera http://www.art <u>Unacceptable</u> <u>Stability (colle</u> 70°C: 1 year	om urine. First morning urine is preferred. <u>paration:</u> Transfer 1 mL urine to an ARUP Standard Transport Tube and freeze immed <u>port Temperature:</u> CRITICAL FROZEN (preferred). Refrigerated specimens are acce- start of collection. ical information is needed for appropriate interpretation. Additional required informat up), drug therapy, and family history. Biochemical Genetics Patient History Form is a uplab.com/patienthistory or by contacting ARUP Client Services. <u>Conditions:</u> Samples exposed to more than two freeze/thaw cycles. <u>section to initiation of testing):</u> Ambient: Unacceptable; Refrigerated: 24 hours; Frozen	iately. (Min: 0.3 mL) ptable for testing if frozen within ion includes age, gender, diet available on the ARUP Web site at at -20°C: 1 week; Frozen at -

Reference Interval:

Age	Pipecolic Acid	Total AASA-P6C
0 - 30 days	Less than or equal to19.6 mmol/mol creatinine	Less than or equal to 18.7 mmol/mol creatinine
1 - 5 months	Less than or equal to 12.1 mmol/mol creatinine	Less than or equal to 13.4 mmol/mol creatinine
6 - 11 months	Less than or equal to7.2 mmol/mol creatinine	Less than or equal to 5.3 mmol/mol creatinine
Greater than 1 Year	Less than or equal to 1.2 mmol/mol creatinine	Less than or equal to 3.1 mmol/mol creatinine

Interpretive Data: Refer to report.

See Compliance Statement B: www.aruplab.com/CS

CPT Code(s): 82542

New York DOH approval pending. Call for status update.

HOT LINE NOTE: Refer to the Test Mix Addendum for interface build information.

0040270 Red Blood Cell Count

Keu bioou Cell Co

Ö

Time Sensitive

Specimen Required: Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 48 hours; Frozen: Unacceptable

Reference Interval: Effective May 16, 2016

Age	0 days	1-6 days	7-13 days	14-29 days	30-60 days	61-90 days	91-180 days	6-23 months	2-5 years	6-11 years	12-17 years	18 years and older
Male (M/µL)	3.9-5.5	4.0-6.6	3.9-6.3	3.6-6.2	3.0-5.4	2.7-4.9	3.1-4.5	3.7-5.3	3.9-5.3	4.0-5.2	4.5-5.3	4.7-6.14
Female (M/µL)	3.9-5.5	4.0-6.6	3.9-6.3	3.6-6.2	3.0-5.4	2.7-4.9	3.1-4.5	3.7-5.3	3.9-5.3	4.0-5.2	4.1-5.1	4.08-5.47

RBC



CHR

0040263 Reticulocyte, Hemoglobin Panel



Time Sensitive

Specimen Required: Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 48 hours; Frozen: Unacceptable

Reference Interval: Effective May 16, 2016

Test Number	Components	Reference I	nterval									
0040022	Reticulocytes, Percent and Number											
		Age	0-13 days	14 days and older								
		Male (K/µL)	39.6-137.5	47-152								
		Female (K/µL)	39.6-137.5	47-127								
		%	2.7-6.6%	1.0-2.6%								
	Cellular Hemoglobin											
		Age	0-180 days	6-23 months	2-5 years	6-11 years	12-17 years	18 years and older				
		Male (pg)	27.6-38.7	28.7-35.7	27.7-37.8	32.4-37.6	30.3-40.4	27.9-37.0				
		Female (pg)	29.2-37.5	30.1-35.7	29.3-37.3	30.4-39.7	29.9-38.4	27.9-37.0				
	Immature Reticulocyte Fraction											
		Age	0-3 days	4-30 days	31-60 days	61-180 days	6-23 months	2-5 years	6-11 years	12-17 years	18 and older	
		%	30.5-35.1	14.5-24.6	19.1-28.9	13.4-23.3	11.4-25.8	8.4-21.7	8.9-24.1	9-18.7	2.9-15.5	
						•		•				

HOT LINE NOTE: There is a component change associated with this test. Add component 2013368, Immature Reticulocyte Fraction



RETICULOCY

S100B CSF

0040022 Reticulocytes, Percent and Number

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Time Sensitive

Specimen Required: Stability (collection to initiation of testing): Ambient: 24 hours; Refrigerated: 48 hours; Frozen: Unacceptable

Reference Interval: Effective May 16, 2016

Test Number	Components	Reference Interval				
	Reticulocytes Percent					
		Age	0-13 days	14 days and older		
		%	2.7-6.6%	1.0-2.6%		
	Reticulocyte Number					
		Age	0-13 days	14 days and older		
		Male (K/µL)	39.6-137.5	47-152		
		Female (K/µL)	39.6-137.5	47-127		

New Test 2013358 S100B, CSF Available April 18, 2016 \$ \$

Methodology:Quantitative Enzyme-Linked Immunosorbent AssayPerformed:TueReported:1-8 days

Specimen Required: Collect: CSF.

<u>Specimen Preparation:</u> Transport 1 mL CSF in an ARUP Standard Transport Tube. (Min. 0.5 mL) <u>Storage/Transport Temperature:</u> Frozen. <u>Unacceptable Conditions:</u> Grossly hemolyzed specimens. <u>Stability (collection to initiation of testing):</u> Ambient: 4 hours; Refrigerated: 1 week; Frozen: 1 month

Reference Interval: 0-690 ng/L

Interpretive Data: This assay is performed using the CanAg S100 Enzyme Immunoassay. Results obtained with different assay methods or kits cannot be used interchangeably.

See Compliance Statement D: www.aruplab.com/CS

CPT Code(s): 83520

New York DOH approval pending. Call for status update.



New Test 2013251 Available April 18, 2016 18

STAT6 by Immunohistochemistry

STAT6 IHC

Methodology: Immunohistochemistry Performed: Mon-Fri Reported: 1-3 days

Specimen Required: Collect: Tissue.

Specimen Preparation: Formalin fix (10 percent neutral buffered formalin) and paraffin embed specimen (cells must be prepared into a cellblock). Protect paraffin block and/or slides from excessive heat. Transport tissue block or 5 unstained (3- to 5-micron thick sections), positively charged slides in a tissue transport kit (ARUP supply #47808 recommended but not required) available online through eSupply using ARUP Connect or contact ARUP Client Services at (800) 522-2787. (Min: 2 slides) If sending precut slides, do not oven bake.

<u>Storage/Transport Temperature:</u> Room temperature. Also acceptable: Refrigerated. Ship in cooled container during summer months. <u>Unacceptable Conditions:</u> Specimens submitted with non-representative tissue type. Depleted specimens. <u>Stability (collection to initiation of testing)</u>: Ambient: Indefinitely, Refrigerated: Indefinitely, Frozen: Unacceptable

Note: All stains will be handled as "Stain and Return" unless a consultation is requested. To request a consultation, submit the pathology report, all associated case materials (clinical history, blocks, slides, etc.), and the Anatomic Pathology requisition form (#32960) in place of the Immunohistochemistry Stain Form.

CPT Code(s): 88342

New York DOH approval pending. Call for status update.



0050725 Streptococcus pneumoniae Antibodies, IgG (14 Serotypes)

PNEUMO AB

Interpretive Data: A pre- and post-vaccination comparison is required to adequately assess the humoral immune response to Prevnar 7 (P7), Prevnar 13 (P13), and/or Pneumovax 23 (PNX) *Streptococcus pneumoniae* vaccines. Pre-vaccination samples should be collected prior to vaccine administration. Post-vaccination samples should be obtained at least 4 weeks after immunization. Testing of post-vaccination samples alone will provide only general immune status of the individual to various pneumococcal serotypes.

In the case of pure polysaccharide vaccine, indication of immune system competence is further delineated as an adequate response to at least 50 percent of the serotypes in the vaccine challenge for those 2-5 years of age and to at least 70 percent of the serotypes in the vaccine challenge for those 6-65 years of age. Individual immune response may vary based on age, past exposure, immunocompetence, and pneumococcal serotype.

Responder Status	Antibody Ratio
Non-Responder	Less than 2-fold
Weak Responder	2-fold to 4-fold
Good Responder	Greater than 4-fold

A response to 50-70 percent or more of the serotypes in the vaccine challenge is considered a normal humoral response¹. Antibody concentration greater than $1.0 - 1.3 \,\mu$ g/mL is generally considered long-term protection².

References:

- 1. Daly TM, Pickering JW, Zhang X, Prince HE, Hill HR. Multilaboratory assessment of threshold versus fold-change algorithms for minimizing analytical variability in multiplexed pneumococcal IgG measurements. *Clin Vaccine Immunol.* 2014;21(7):982-8.
- 2. Daly TM, Hill HR. Use and Clinical Interpretation of Pneumococcal Antibody Measurements in the Evaluation of Humoral Immune Function. *Clin Vaccine Immunol.* 2015;22(2):148-152.

See Compliance Statement B: www.aruplab.com/CS

HOT LINE NOTE: Remove information found in the Note field.

HOT LINE NOTE: There is a clinically significant charting name change associated with this test. Change the charting name of component 0050706 from Pneumococcal Serotype 1 IgG to Pneumo serotype 1 IgG (P13,PNX) Change the charting name of component 0050709 from Pneumococcal Serotype 4* IgG to Pneumo serotype 4 IgG (P7,P13,PNX) Change the charting name of component 0050709 from Pneumococcal Serotype 5 IgG to Pneumo serotype 5 IgG (P13,PNX) Change the charting name of component 0050713 from Pneumococcal Serotype 6B* IgG to Pneumo serotype 6B IgG (P7,P13,PNX) Change the charting name of component 0050715 from Pneumococcal Serotype 3 IgG to Pneumo serotype 3 IgG (P13,PNX) Change the charting name of component 0050716 from Pneumococcal Serotype 7F IgG to Pneumo serotype 7F IgG (P13,PNX) Change the charting name of component 0050717 from Pneumococcal Serotype 9N IgG to Pneumo serotype 7F IgG (P13,PNX) Change the charting name of component 0050717 from Pneumococcal Serotype 9N IgG to Pneumo serotype 9N IgG (PNX) Change the charting name of component 0050712 from Pneumococcal Serotype 8 IgG to Pneumo serotype 8 IgG (P7,P13,PNX) Change the charting name of component 0050721 from Pneumococcal Serotype 8 IgG to Pneumo serotype 8 IgG (PNX) Change the charting name of component 0050722 from Pneumococcal Serotype 8 IgG to Pneumo serotype 9V IgG (P7,P13,PNX) Change the charting name of component 0050723 from Pneumococcal Serotype 12F IgG to Pneumo serotype 12F IgG (PNX) Change the charting name of component 0050724 from Pneumococcal Serotype 12F IgG to Pneumo serotype 12F IgG (P7,P13,PNX) Change the charting name of component 0050724 from Pneumococcal Serotype 18C* IgG to Pneumo serotype 18C IgG (P7,P13,PNX) Change the charting name of component 0050726 from Pneumococcal Serotype 19F* IgG to Pneumo serotype 19F IgG (P7,P13,PNX) Change the charting name of component 0050727 from Pneumococcal Serotype 19F* IgG to Pneumo serotype 19F IgG (P7,P13,PNX)



2005779 Streptococcus pneumoniae Antibodies, IgG (23 Serotypes)

PNEUMO 23

Interpretive Data: A pre- and post-vaccination comparison is required to adequately assess the humoral immune response to Prevnar 7 (P7), Prevnar 13 (P13), and/or Pneumovax 23 (PNX) Streptococcus pneumoniae vaccines. Pre-vaccination samples should be collected prior to vaccine administration. Post-vaccination samples should be obtained at least 4 weeks after immunization. Testing of post-vaccination samples alone will provide only general immune status of the individual to various pneumococcal serotypes.

In the case of pure polysaccharide vaccine, indication of immune system competence is further delineated as an adequate response to at least 50 percent of the serotypes in the vaccine challenge for those 2-5 years of age and to at least 70 percent of the serotypes in the vaccine challenge for those 6-65 years of age. Individual immune response may vary based on age, past exposure, immunocompetence, and pneumococcal serotype.

Responder Status	Antibody Ratio
Non-Responder	Less than 2-fold
Weak Responder	2-fold to 4-fold
Good Responder	Greater than 4-fold

A response to 50-70 percent or more of the serotypes in the vaccine challenge is considered a normal humoral response¹. Antibody concentration greater than $1.0 - 1.3 \,\mu$ g/mL is generally considered long-term protection².

References:

- Daly TM, Pickering JW, Zhang X, Prince HE, Hill HR. Multilaboratory assessment of threshold versus fold-change algorithms for minimizing analytical variability in multiplexed pneumococcal IgG measurements. Clin Vaccine Immunol. 2014;21(7):982-8.
- 2. Daly TM, Hill HR. Use and Clinical Interpretation of Pneumococcal Antibody Measurements in the Evaluation of Humoral Immune Function. Clin Vaccine Immunol. 2015;22(2):148-152.

See Compliance Statement B: www.aruplab.com/CS

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2008919 *Streptococcus pneumoniae* Antibodies, IgG (9 Serotypes)

PNEUMO 9

Interpretive Data: A pre- and post-vaccination comparison is required to adequately assess the humoral immune response to Prevnar 7 (P7), Prevnar 13 (P13), and/or Pneumovax 23 (PNX) *Streptococcus pneumoniae* vaccines. Pre-vaccination samples should be collected prior to vaccine administration. Post-vaccination samples should be obtained at least 4 weeks after immunization. Testing of post-vaccination samples alone will provide only general immune status of the individual to various pneumococcal serotypes.

In the case of pure polysaccharide vaccine, indication of immune system competence is further delineated as an adequate response to at least 50 percent of the serotypes in the vaccine challenge for those 2-5 years of age and to at least 70 percent of the serotypes in the vaccine challenge for those 6-65 years of age. Individual immune response may vary based on age, past exposure, immunocompetence, and pneumococcal serotype.

Responder Status	Antibody Ratio
Non-Responder	Less than 2-fold
Weak Responder	2-fold to 4-fold
Good Responder	Greater than 4-fold

A response to 50-70 percent or more of the serotypes in the vaccine challenge is considered a normal humoral response¹. Antibody concentration greater than $1.0 - 1.3 \,\mu$ g/mL is generally considered long-term protection².

References:

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- 2. Daly TM, Hill HR. Use and Clinical Interpretation of Pneumococcal Antibody Measurements in the Evaluation of Humoral Immune Function. *Clin Vaccine Immunol*. 2015;22(2):148-152.

See Compliance Statement B: www.aruplab.com/CS

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New Test	<u>2013325</u>	Systemic Scleroderma Comprehensive Panel	SCL COMP
Available July	5, 2016		
Methodology:	Semi-Quantita Quantitative E	tive Immunoblot/Semi-Quantitative Indirect Fluorescent Antibody/Semi-Quantitanzyme-Linked Immunosorbent Assay	tive Multiplex Bead Assay/Semi-
Performed:	Tue		
Reported:	1-8 days		
Specimen Require	ed: Collect: Serum	Separator Tube (SST).	
	Specimen Prep	aration: Separate from cells ASAP or within 2 hours of collection. Transfer 3 mL	serum to an ARUP Standard
	Transport Tub	e. (Min: 1.5 mL)	
	Storage/Transp	ort Temperature: Refrigerated. Also acceptable: Room temperature or frozen.	
	Unacceptable	Conditions: Hemolyzed, hyperlipemic, icteric, heat-treated or contaminated specin	nens

Stability (collection to initiation of testing): Ambient: 48 hours; Refrigerated: 2 weeks; Frozen: 1 year

Reference Interval:

Test Number	Components	Reference Interval
0050599	Scleroderma (Scl-70)	29 AU/mL or less: Negative
	(ENA) Antibody, IgG	30-40 AU/mL: Equivocal
		41 AU/mL or greater: Positive
0050470	Ribonucleic Protein (U1)	29 AU/mL or less: Negative
	(ENA) Ab, IgG	30-40 AU/mL: Equivocal
		41 AU/mL or greater: Positive
0050714	Centromere Ab, IgG	29 AU/mL or less: Negative
		30-40 AU/mL: Equivocal
		41 AU/mL or greater: Positive
0050639	Anti-Nuclear Antibody	Less than 1:40
	(ANA), IgG by IFA	
2012173	Fibrillarin (U3 RNP) Ab,	5 Units or less: Negative
	IgG	6-10 Units: Equivocal
		11 Units or greater: Positive
2003040	PM/Scl 100 Antibody,	5 Units or less: Negative
	IgG	6-10 Units: Equivocal
		11 Units or greater: Positive
2001601	RNA Polymerase III	19 Units or less: Negative
	Antibody, IgG	20-39 Units: Weak Positive
		40-80 Units: Positive
		81 Units or greater: Strong Positive

Interpretive Data: Refer to report.

Note: Panel includes: Anti-Nuclear Ab (ANA) Titer, Anti-Nuclear Ab (ANA) Pattern, Anti-Scl-70, Anti-RNA Polymerase III Ab, Anti-Centromere Ab, Anti-U1 RNP Ab, Anti-Fibrillarin (U3 RNP), Anti-PM/Scl Ab.

CPT Code(s): 86039, 86235 x4, 83516 x2

New York DOH Approved.



New Test Available May	2013275Tartrate-Resistant Acid Phosphatase, Cytochemical Stain OnlyTRAP SO16, 2016
Ō	Time Sensitive
Methodology: Performed: Reported:	Cytochemical Stain Mon-Fri 2-5 days
Specimen Require	 collect: Lavender (EDTA), Green (Lithium Heparin), or Green (Sodium Heparin). Also acceptable: Heparinized Bone Marrow Aspirate. <u>Specimen Preparation:</u> Transport 6 unfixed, air-dried, and unstained push smears AND 5 mL whole blood (smears should be made from the blood submitted). OR transport 6 unfixed, air-dried, and unstained bone marrow aspirate smears. Protect from light. <u>Storage/Transport Temperature:</u> Room temperature. Blood specimens must be received within 24 hours of collection; testing must be performed within 48 hours of collection. <u>Unacceptable Conditions:</u> Peripheral blood smears older than one year. Anticoagulated blood or bone marrow sent without well-prepared unfixed smears if greater than 48 hours. <u>Stability (collection to initiation of testing)</u>: Ambient: 24 hours; Refrigerated: Unacceptable; Frozen: Unacceptable
Note: Further info https://www.youtu	ormation on how to make an adequate slide, in the form of an instructional video, can be found at: be.com/watch?v=ca3NwrlpS40&feature=youtube
CPT Code(s):	88319
HOT LINE NO	TE: Refer to the Test Mix Addendum for interface build information.

New Test	2013335 Thrombopoietin (TPO), Serum	THROMBO S						
Available April 1	8, 2016							
Mathadala any								
Methodology:	Quantitative Immunoassay							
Performed:	Varies							
Reported:	6-13 days							
Specimen Required:	Collect: Plain Red. Also acceptable: Serum Separator Tube (SST)							
	Specimen Preparation: Transfer 1 mL serum to an ARUP Standard Transport Tube. (M	Min: 0.5 mL)						
	Storage/Transport Temperature: Frozen.							
	Stability (collection to initiation of testing): Ambient: Unacceptable; Refrigerated: Un	acceptable; Frozen: 1 month						
		•						
Reference Interval: By Report								

CPT Code(s): 83520

New York DOH Approved.

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New Test	2013290 Tropheryma whipplei PCR	TWHIPPCR						
Available April	18, 2016							
Methodology: Performed: Reported:	Qualitative Polymerase Chain Reaction Tue, Fri 2-5 days							
Specimen Required	I: <u>Collect:</u> Lavender (EDTA), pink (K ₂ EDTA), Serum Separator Tube (SST), or CSF. <u>Specimen Preparation</u> : Transfer 1 mL serum, plasma, whole blood, or CSF to a sterile container. (Min: 0.5 mL <u>Storage/Transport Temperature: Frozen.</u> <u>Remarks</u> : Specimen source required. <u>Unacceptable Conditions</u> : Heparinized specimens. <u>Stability (collection to initiation of testing)</u> : Ambient: 8 hours; Refrigerated: 5 days; Frozen: 2 weeks	.)						
Interpretive Data	: See Compliance Statement B: www.aruplab.com/CS							
CPT Code(s):	87798							
New York DOH app	roval pending. Call for status update.							
HOT LINE NOT	E: Refer to the Test Mix Addendum for interface build information.							
2005413	Urticaria-Inducing Activity	UIA						
Specimen Required	Stability (collection to initiation of testing): After separation from cells: Ambient: Unacceptable; Refrigerated: year (avoid repeated freeze/thaw cycles)	Unacceptable; Frozen: 1						
0080380	Vitamin C (Ascorbic Acid), Plasma	VIT C						
Specimen Required	: <u>Unacceptable Conditions</u> : EDTA plasma, whole blood, or body fluids. Grossly hemolyzed specimens.							
0040320	White Blood Cell Count	WBC						
Ö	Fime Sensitive							

Specimen Required: Collect: Lavender (EDTA) or Pink (K₂ EDTA).

<u>Specimen Preparation:</u> Mix specimens thoroughly. Transport 5 mL whole blood. (Min: 0.25 mL) <u>Storage/Transport Temperature:</u> Refrigerated. <u>Stability (collection to initiation of testing):</u> Ambient: 24 hours; Refrigerated: 48 hours; Frozen: Unacceptable

Reference Interval: Effective May 16, 2016

Age	0 days	1-6 days	7-13 days	14-29 days	30-60 days	2-11 months	1-3 years	4-5 years	6-7 years	8-13 years	14-17 years	18 years and older
Male (K/µL)	9-30	9-34	5-21	5-20	5-19.5	5.5-17	6-17	5.5-15.5	5-14.5	4.5-13.5	4.5-13	4.3-11.3
Female (K/µL)	9-30	9-34	5-21	5-20	5-19.5	5.5-17	6-17	5.5-15.5	5-14.5	4.5-13.5	4.5-13	4.3-11.3



The following will be discontinued from ARUP's test menu on July 5, 2016. Replacement test options are supplied if applicable.

Test Number	Test Name	Refer To Replacement
0055566	Apolipoprotein E (APOE) 2 Mutations, Cardiovascular Risk	APOE CR (2013337)
0055691	BIRC2-MALT1 (API2-MALT1) Translocation, t(11;18) by RT-PCR	Chromosome FISH, Interphase (2002298)
0051434	Bloom Syndrome (BLM) 1 Mutation, Fetal	
0051454	Canavan Disease (ASPA) 4 Mutations, Fetal	
0021021	Carotenes, Fractionated, Plasma or Serum	
0055283	Cysticercosis Antibody, IgG by Western Blot	
0055282	Cysticercosis Antibody, IgG by Western Blot (CSF)	
<u>2008920</u>	Cytochrome P450 Pain Management Panel, CYP2D6, CYP2C9, CYP2C19 - Common Variants	Cytochrome P450 Genotype Panel (2013098)
0091589	Darvocet, Urine	
0051464	Dysautonomia, Familial (IKBKAP) 2 Mutations, Fetal	
0051469	Fanconi Anemia, Group C (FANCC) 2 Mutations, Fetal	
0030140	Fibrin/Fibrinogen Degradation Split Products	FDP Plasma (2006491)
0091509	Formic Acid, Urine	
0051439	Gaucher Disease (GBA) 8 Mutations, Fetal	
0091353	Glyburide Quantitative, Serum or Plasma	
<u>0050567</u>	Inflammatory Bowel Disease Differentiation Profile	Inflammatory Bowel Disease Differentiation Panel (2013270)
<u>0091530</u>	Inhalants Panel, Solvents, Serum or Plasma	
0051449	Mucolipidosis, Type IV (MCOLN1) 2 Mutations, Fetal	
2012535	Nerve Fiber Density Analysis, Intraepidermal	
0051459	Niemann-Pick, Type A (SMPD1) 4 Mutations, Fetal	
2008868	Nonalcoholic steatohepatitis (NASH) FibroSURE	
2006462	Scleroderma Antibodies Panel	Systemic Scleroderma Comprehensive Panel (2013325)
0051429	Tay-Sachs Disease (HEXA) 7 Mutations, Fetal	
2011025	Tropheryma whipplei Detection by PCR, Blood	Tropheryma whipplei PCR (2013290)
2008116	Urine Culture, Invasive Collection	Urine Culture (0060131)
<u>2004175</u>	Vascular Endothelial Growth Factor C (VEGF-C) by Immunohistochemistry	
2011127	Zolpidem and Metabolites Quantitative, Urine	Zolpidem, Urine, Quantitative (2012319)